

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re Application of:

HEATH, *et al.*

Serial No.: 08/699,716

Filed: 27 August 1996

For: RECOMBINANT F1-V PLAGUE VACCINE



Art Unit: 1645

Examiner: Duffy, Patricia Ann

Atty. Dckt: 003/029/SAP

AFFIDAVIT OF GEORGE W. ANDERSON, JR.

1. I, George W. Anderson, Jr., an inventor of the above-referenced application and resident of Smithsburg, MD, declare the following:
2. My curriculum vitae is attached.
3. Arthur M. Friedlander, David G. Heath, Susan L. Welkos and I are joint inventors of the subject matter disclosed in the above-referenced application.
4. From [redacted date which is before 13 March 1996] to February 1998, I conducted research and development on a plague vaccine comprising a F1-V fusion protein as an immunogen as part of the Army Plague Vaccine Group.
5. Before about [redacted date which is before 13 March 1996], I obtained alhydrogel F1-V partial preparations from David G. Heath.
6. In my laboratory notebook I usually referred to the F1-V partial as "F1-V".
7. On [redacted date which is before 13 March 1996], I began mouse challenge studies with the F1-V partial preparations I obtained from David G. Heath. The experimental protocol for the challenge studies is provided in my notebook #3598 on page 123. See Exhibit GA1.
8. The results dated [redacted date which is before 13 March 1996] for the challenge studies using *Yersinia pestis* strain CO92, which is F1⁺ strain, with mice immunized alhydrogel F1-V partial preparations and controls with are provided in my notebook #3598 on pages 125-126. See Exhibit GA2.
9. The results dated [redacted date which is before 13 March 1996] for the challenge studies using *Yersinia pestis* strain C12, which is F1⁻ strain, with mice immunized with alhydrogel F1-V partial preparations and controls with are provided in my notebook #3598 on pages 127-130. See Exhibit GA3.
10. On [redacted date which is before 13 March 1996], I wrote in my notebook #3598 on page 132, that the data on page 131 of my notebook and the mouse challenge studies are the first direct evidence that the F1-V fusion (F1-V partial) can induce an immune response to both F1 and V protein. David G. Heath witnessed this page and the results to which it references. See Exhibit GA4.
11. Exhibit AF3 (GA5) is an excerpt of my notebook #3598.
12. On [redacted date which is before 13 March 1996], I gave David G. Heath the protocol for formulating the F1-V whole vaccine preparations for mouse challenge assays. See Exhibit

DH16 (GA6).

13. In my laboratory notebook, I usually referred to the F1-V whole as "F1-WV".
14. Before about [redacted date which is before 13 March 1996], I obtained alhydrogel F1-V whole preparations from David G. Heath and began conducting the mouse challenge studies. See Exhibit GA7.
15. The results of the challenge studies dated [redacted date which is before 13 March 1996] using *Yersinia pestis* strain C12 or CO92 with mice immunized with alhydrogel F1-V partial preparations and controls are provided in my notebook #3739 on pages 60-63. The results show that F1-V whole confer a protective immune response against both F⁺ and F⁻ *Yersinia pestis* strains. See Exhibit GA8.
16. From about [redacted date which is before 13 March 1996] to 27 August 1996, I conducted further experiments to determine the efficacy of the F1-V fusion proteins and to determine whether any refinements could be made, such as the following:
 - a. On [redacted date which is before 13 March 1996], I conducted a mouse challenge study examining the long term efficacy of F1-V whole which is documented in my notebook #3739, page 75. See Exhibit GA9. The results dated [redacted date which is before 13 March 1996] are provided in my notebook #3739, page 85. See Exhibit GA10.
 - b. On [redacted date which is before 13 March 1996], I conducted a mouse challenge study examining the range of Al concentration which maintains an adequate adjuvant response which is documented in my notebook #3739, page 88. See Exhibit GA11. The alhydrogel F1-V whole preparations were obtained from David G. Heath. I copied the notebook pages from David G. Heath's notebook and inserted in my notebook. See Exhibit GA12. The results dated [redacted date which is before 13 March 1996] are found in my notebook #3739, pages 104-107. See Exhibit GA13.
 - c. Exhibit GA14 shows experimental data from my notebook #3739 which evidence that from [redacted date which is before 13 March 1996] to 23 February 1996, I conducted various mouse challenge studies with F1-V whole.
 - d. On 3 April 1996, I obtained the results for ELISA assays of serum obtained from mice immunized with F1-V whole to determine if the sera still contained antibodies against F1 antigen and V antigen as evidenced in my notebook #3739, page 122. See Exhibit GA15.
 - e. On about 15 May 1996, I conducted a study with mice vaccinated with different amounts of F1-V whole protein. The mice were challenged subcutaneously and by aerosol with *Y. pestis*, C092 or C12. See Exhibit GA16.
 - f. On about 28 June 1996, I conducted a study with mice vaccinated with either F1-V whole or a mixture of F1 + V or Plague USP vaccine. The mice were challenged by aerosol with *Y. pestis* C092. See pages 134 and 137 of my notebook #3739, Exhibit GA17.
 - g. On about 5 July 1996, I conducted a study with mice vaccinated with either F1-V whole or F1 + V or Plague USP vaccine. The mice were challenged

subcutaneously and by aerosol with *Y. pestis*, C092 or C12. See pages 135-136 of my notebook #3739, Exhibit GA18.

17. For all the challenge studies referenced herein, I obtained most of the *Yersinia pestis* strains C12 and CO92 from Susan L. Welkos.
18. On 15 February 1996, I presented the work summarized in David G. Heath's Abstract 17. See Exhibit DH19 (GA19).
19. I left the Army Plague Vaccine Group on 26 February 1998 when I retired from the U.S. Army.
20. I have reviewed and analyzed the Titball patent and the three priority documents, UK 9505059, UK 9518946, and UK 9524825, and PCT/GB96/00571.
21. It is my opinion that prior to 13 March 1996, the filing date of PCT/GB96/00571, the inventors of the Titball patent had not conceived and/or reduced to practice a plague vaccine comprising purified F1 antigen fused to all or part of V antigen as nowhere do UK 9505059, UK 9518946, and UK 9524825 disclose isolating or purifying a protein comprising F1 antigen fused to all or part of V antigen from the host cell and other cellular components and/or administering a purified protein comprising F1 antigen fused to all or part of V antigen to a subject.
 - a. In fact, UK 9518946 is the first disclosure indicating a genetic vaccine or how a host organism may be transfected with DNA for F1 antigen and V antigen to result in a live vaccine, i.e. an attenuated host organism (such as *Salmonella*) which produces the antigen when administered to a subject.
 - b. As described in UK 9518946, the genetic vaccine or the live vaccine is administered to a subject such that the protein/antigen of interest is then produced in the subject.
 - c. UK 9518946 does not describe isolating the protein/antigen of interest from the host organism and purifying the protein/antigen of interest from other cellular components prior to administration to a subject.
 - d. The genetic vaccine or live vaccine described in UK 9518946 is not a purified protein comprising F1 antigen fused to all or part of V antigen which is isolated and purified from cells and other cellular components as claimed in the above-referenced application.
22. I have reviewed and analyzed the experiments and data of the Army Plague Vaccine Group and it is my opinion that the Army Plague Vaccine Group:
 - a. Conceived of a fusion protein comprising F1 antigen fused to part of V by at least [redacted date which is before 13 March 1996].
 - b. Conceived of a fusion protein comprising F1 antigen fused to all of V by at least [redacted date which is before 13 March 1996].
 - c. Conceived of and reduced to practice a purified fusion protein comprising F1 antigen fused to part of V by at least [redacted date which is before 13 March 1996].
 - d. Conceived of and reduced to practice a purified fusion protein comprising F1

antigen fused to all of V by at least [redacted date which is before 13 March 1996].

- e. Conceived of and reduced to practice a vaccine against plague comprising a purified fusion protein comprising F1 antigen fused to part of V by at least [redacted date which is before 13 March 1996].
- f. Conceived of and reduced to practice a vaccine against plague comprising a purified fusion protein comprising F1 antigen fused to all of V by at least [redacted date which is before 13 March 1996].

23. I declare that all statements made herein of my own knowledge are believed to be true, and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under §1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.



Date: 14 March 2007

George W. Anderson, Jr.

CURRICULUM VITAE

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PROFESSIONAL EXPERIENCE:

2004-Present Senior Principal Engineer. Scientific responsibilities for the Operations and Sustainment of a Defense Threat Reduction sponsored multi-country epidemiological surveillance system and collaborative biological research program in the former Soviet Union.

2003-2004 Principal Advisor. Midwest Research Institute. General scientific guidance to the company and responsibilities for integrating the capabilities of various company divisions in projects. Continue participation in DTRA projects in the former Soviet Union (Kazakhstan, Uzbekistan and Republic of Georgia).

Senior Program Manager. Department at Southern Research Institute was purchased by Midwest Research Institute. I continued program management of the work in the former Soviet Union until promoted to Principal Advisor.

1998-2003 Director, Medical Countermeasures Department. Southern Research Institute. Responsible for the management, direction, control and review of the departmental research and development programs. Established a Biological Safety Level-3 containment facility for vaccine potency testing of Department of Defense (DoD) Investigational New Product (IND) vaccines (VEE, WEE, EEE, Q fever, Tularemia) in Frederick, MD. Department provided the DoD with expertise in biological defense, biosecurity, biotechnology and

biosafety for DARPA and DTRA projects at former biological weapons facilities in the former Soviet Union (FSU). Some of the projects support Non-proliferation efforts. Projects in the FSU include on-site observations of laboratory work in BSL-2, -3 and BSL-4 containment laboratories. Worked as a consultant to the Department of the Army for the anthrax vaccine production facility in the United States. Conducted audits in BSL-3 laboratories. Immunized with most licensed and IND bio-defense vaccines. Biosafety consultant to U.S. laboratories. Technical spokes person on biosafety for dismantlement of a pilot production facility for biological warfare agents in the United States.

1998-1998 Principal Scientist, SRS Technologies. Perform and manage scientific and technical tasks requiring the assessment of chemical (CW) or biological warfare (BW) programs/capabilities of foreign countries for terrorist groups and the development of recommended measures to curb the proliferation of biological weapons and technologies.

1993-1998 Chief, Pathogenesis and Immunology Branch, U.S. Army Medical Research Institute of Infectious diseases (USAMRIID). Branch Chief responsible for directing research programs directed toward development and production of prophylactic and therapeutic modalities against bacterial diseases of potential biological warfare significance (e.g., anthrax, plague, Q fever, tularemia, glanders). Special project officer for multi-million dollar Good Laboratory Practices project which included facility upgrades, Responsible for the GLP BSL-3 laboratory setup, and project manager for multi-year preclinical project for a supplement to the anthrax vaccine license. Research associate on the clinical protocol for a supplement to the current anthrax vaccine license. Manage technical aspects of a contract for cGMP production of cell banks and recombinant *Bacillus anthracis* PA protein as diagnostic or vaccine component. Involved with nonproliferation activities with former Soviet weapons scientist. First US military officer invited into the BSL-3 containment laboratories at the State Research Center of Applied Microbiology; Obolensk, Russia.

1990-1993 Medical R&D Officer, Science and Technology Center-Europe, Frankfurt, Germany - Responsible for finding biotechnologies, products, and collaboration in Europe, the Middle East, Africa, and the former Soviet Union, which could shorten or negate the need for the R&D cycle in Defense Department laboratories. This work involved extensive travel to scientific conference and Institutes in these geographical areas and technical report writing.

1987-1990	Research Immunologist, U.S. Army Medical Research Institute of Infectious Diseases. Investigations included development of a congenic strain of rats, vaccine efficacy trials against an aerosol exposure, and pathogenesis studies on Rift Valley fever virus. These studies were carried out in a Biosafety Level 3 laboratory.
1983-1987	Graduate student at the Johns Hopkins University, while on active duty, US Army.
1977-1983	Research Scientist, U.S. Army Medical Research Institute of Infectious Diseases (USAMRIID), Fort Detrick, Frederick, Maryland Was responsible for developing and investigating genetically defined animal models for exotic viral and rickettsial diseases.

ACADEMIC APPOINTMENT

Associate Professor, Center for Disaster Preparedness, School of Medicine, University of Alabama

FIELD STUDIES:

1988 Member of a team of two who conducted an epidemiological sero-survey for phleboviral infections at the MRC Laboratory, The Gambia, and at the Institute Pasteur, Senegal, West Africa.

MILITARY SERVICE:

1977-1998 Retirement rank: LTC, U.S. Regular Army Commission, MSC
Army Management Staff College, graduate
Command and General Staff College, graduate

EDUCATION:

Ph.D., 1988 The Johns Hopkins University, Baltimore, Maryland, Viral Immunology. Dissertation: *Viral and Host Determinants of Resistance to Rift Valley Fever in a Rat Model*

M.S., 1977 Florida Institute of Technology, Melbourne, Florida, Biology.

B.S., 1975 Florida Institute of Technology, Melbourne, Florida, Biology.

TEACHING EXPERIENCE:

1996 Mentor, Department of Defense, Science & Engineering Apprentice Program

1989-1990 Served as a thesis committee member for a master's level candidate at Hood College, Frederick, Maryland. Supervised the candidate's research.

1988-1989 Sponsored a Korean ophthalmologist for the 1988-1989 ROK/US Scientist/Engineer Exchange Program in my laboratory to develop a model to study Rift Valley fever ocular sequelae.

1975-1977 Teaching assistantship at Florida Institute of Technology with primary teaching responsibilities for microbiology and biochemistry laboratories.

MEMBERSHIP IN ACADEMIC AND PROFESSIONAL SOCIETIES:

Membership: American Society for Microbiology
Sigma XI Scientific Research Society

HONORS AND AWARDS:

Who's Who Among Students in American Universities – 1974, 1975
Four-year Army ROTC Scholarship
Four-year U.S. Army Long Term Health, Education and Training
Program (Ph.D. scholarship to the Johns Hopkins University)
Blue Key National Honor Fraternity
Sigma XI Scientific Research Society
Scouting: Highest rank – Eagle, Highest honor – Order of the Arrow,
Vigil Member
U.S. Army Army Commendation Medal
 Meritorious Service Award w/2 Oak leaf clusters
 Legion of Merit
Distinguished Professional Achievement Award, Florida Institute of
Technology 2002
Letter of appreciation from Joint Program Office for Biological
Defense for assistance to BioPort Corporation in obtaining
FDA approval for Biological License Agreement for anthrax
vaccine.

PATENTS: U.S. Patent Number 5,320,069, "Small Animal Restraint Device"
Patent Pending Recombinant F1-V Plague Vaccine, filing #08/699,716
18 Dec 96

CONTINUING EDUCATION:

The Regulatory Process and Good Clinical Practices, Technology Management Integration, Inc., 1994

Regulatory Issues in Biotechnology, Univ. MD, 1995
Good Manufacturing Processes for Bioprocesses, Univ. MD, 1996
Quality Control and Quality Assurance of Biotechnology Products,
Univ. MD, 1996
Assay Validation, PDA, 1996
Intro. to GLPs and Auditing, International Center for Health &
Environmental Education, 1996
Writing and Evaluating Standard Operating Procedures for the
Regulatory Environment, International Quality Training, 1996
Intro. FDA Good Laboratory Practices & Documentation Principles,
International Quality Training, 1996
Good Laboratory Practices Regulations for Study Directors,
International Quality Training, 1996
Fundamentals & Concepts of Calibration & Metrology, PDA, 1996
Biotechnology GMP Facility Design, Construction and Validation,
Univ. MD, 1997
Advances in Filtration and Bioseparation Technologies, Pall Ultrafine
Filtration Company, Columbia, MD, 1997
Validation of Biotechnology Processes and Systems, Univ. MD, 1997
Fermentation Microbiology, American Type Culture Collection
Workshop, Rockville, MD, 1997
Fundamentals of D, F and Z Values, PDA, 1997
Basic Principles in Preparation of Sterile Dosage Forms, PDA, 1997
Parenteral Packaging: Rubber, Glass, Plastic and Metal Seals, PDA,
1997
Regulatory Compliance Training, Southern Research Institute, 1998
ISO 9001/Quality system Introductory Training, Southern Research
Institute, 1998
Introductory to Earned Value Seminar, Dynport, LLC Professional
Development, 1998
Positive Pressure Pneumatic BSL-4 Suite Training, State Research
Center of Virology and Biotechnology, "Vector", Russia 2000
Introduction To Aerosol Mechanics I & II, AAAR, 2000
USA-Russia Workshop on International Research Ethics; Institutional
Review Boards and Laboratory Animal Welfare, 2002
Introduction to Laboratory Ventilation and Design, American
Biological Safety Association, 2002
Plant Biosafety, American Biological Safety Association, 2002
Bechtel Safety Leadership Workshop, Bechtel National Inc, 2006
The Transport of diagnostic & Infectious Samples, American
Biological Safety Association, 2006
Biohazard Risk Assessment, American Biological Safety Association,
2006

Certifications:

Transport of diagnostic & Infectious Substances by Air (per ICAO
Technical Instructions & IATA DGR) valid until 15 Oct 2008

REVIEWER/CONSULTANT:

U.S. Army In-house laboratory independent Research (ILIR)
proposals, 1993-1998
U.S. Army Broad Agency Announcement proposals, 1993-1998
Experts Contact for database to assist Biological Arms Control Treaty
Office (BACTO), 1997-1998
Nonproliferation Programs IntraAgency Roundtable, 1997-1998
U.S.-Uzbek Collaborative Biotechnology Grants Program, 2000
Consultant for DoD at BioPort Corporation (anthrax vaccine
production facility), 2000-2002

PROFESSIONAL PUBLICATIONS:

1. **Anderson, G.W., Jr.**, and J.V. Osterman. 1980. Host defenses in experimental Rickettsialpox: Genetics of natural resistance to infection. *Infect. Immun.* 28: 132-136.
2. **Anderson, G.W., Jr.**, and J.V. Osterman. 1980. Host defenses in Rickettsialpox: Resistance of C3H mouse sublines. *Acta. Virol.* 24: 294-296.
3. Peters, C.J., and **G.W. Anderson, Jr.** 1981. Pathogenesis of Rift Valley fever, pp. 21-41. In Contributions to Epidemiology and Statistics, vol. 3. (N. Goldblum, T.A. Swartz, and M.A. Kingberg, eds.). S. Karger, Basel.
4. **Anderson, G.W., Jr.**, T.W. Slone, Jr., and C.J. Peters. 1987. Pathogenesis of Rift Valley fever virus (RVFV) in inbred rats. *Microb. Pathogen.* 2: 283-293.
5. **Anderson, G.W., Jr.**, and J.F. Smith 1987. Immunoelectron microscopy of Rift Valley fever viral morphogenesis in primary rat hepatocytes. *Virology.* 161: 91-100.
6. **Anderson, G.W., Jr.**, and C.J. Peters. 1988. Viral determinants of virulence for Rift Valley fever (RVF) in rats. *Microb. Pathogen.* 5: 241-250.
7. **Anderson, G.W., Jr.**, T.W. Slone, Jr., and C.J. Peters. 1988. The gerbil, *Meriones unguiculatus*. A model for Rift Valley fever viral encephalitis. *Archives Virology* 102: 187-196.
8. **Anderson, G.W., Jr.**, J.-F. Saluzzo, T.G. Ksiazek, J.F. Smith, W. Ennis, D. Thureen, C.J. Peters, and J.P. Digoutte. 1989. Comparison of in vitro and in vivo systems for propagation of Rift Valley fever virus from clinical specimens. *Res. Virol.* 140:129-138.
9. Saluzzo, J.F., **G.W. Anderson, Jr.**, L.A. Hodgson, J.P. Digoutte, and J.F. Smith. 1989. Antigenic and biological properties of Rift Valley fever virus isolated during the 1987 Mauritanian epidemic. *Res. Virol.* 140:155-164.
10. Peters, C.J., C.-T. Liu, **G.W. Anderson, Jr.**, J.C. Morrill, and P.B. Jahrling. 1989. Pathogenesis of viral hemorrhagic fevers: Rift Valley fever and Lassa fever contrasted. *Rev. Infect. Dis.* 11 (Suppl. 4): 5743-5749.
11. Saluzzo, J.F., **G.W. Anderson, Jr.**, J.F. Smith, D. Fontenille, and P. Coulanges. 1989. Biological and antigenic relationship between Rift Valley fever virus strains isolated in Egypt and Madagascar. *Trans. R. Soc. Trop. Med. Hyg.* 83:701.
12. Solow, R., K. Mereish, **G.W. Anderson, Jr.**, and J. Hewetson. 1990, Effect of microcystin-LR on cultured rat endothelial cells. *Med. Sci. Res.* 18:241-244.

13. **Anderson, G.W., Jr.**, M.V. Slayter, W. Hall, and C.J. Peters. 1990. Pathogenesis of a phleboviral infection (Punta Toro virus) in Golden Syrian hamsters. *Arch. Virol.* 114: 203-212.
14. **Anderson, G.W., Jr.**, J.O. Lee, A.O. Anderson, N. Powell, J.A. Mangiafico, and G. Meadors. 1991. Efficacy of a Rift Valley fever virus vaccine against an aerosol infection in rats. *Vaccine*. 9: 710-714.
15. **Anderson, G.W., Jr.**, J.A. Rosebrock, A.J. Johnson, G.B. Jennings, and C.J. Peters. 1991. Infection of inbred rats with Rift Valley Fever virus: Development of a congenic resistant strain and observations on age-dependence of resistance. *Am. J. Trop. Med. Hyg.* 44(5): 475-480.
16. **Anderson, G.W., Jr.**, W.B. Lawrence, J-O Lee, and M. Young. 1991. A restraint for ophthalmic examination of unanesthetized rats. Note. *Laboratory Animal Science*. 41(3): 288-290.
17. Friedlander, A.M., S.L. Welkos, P.L. Worsham, G.P. Andrews, D.G. Heath, **G.W. Anderson, Jr.**, M.L.M. Pitt, J. Estep, and K. Davis. 1995. Relationship between virulence and Immunity as revealed in recent studies of the F1 capsule of *Yersinia pestis*. *Clinical Infectious Diseases*. 21(Suppl 2):S178-81.
18. Andrews, G.P., D.G. Heath, **G.W. Anderson, Jr.**, S.L. Welkos, and A.M. Friedlander. 1996. Fraction 1 capsular antigen (F1) purification from *Yersinia pestis* CO92 and an *Escherichia coli* recombinant strain and efficacy against lethal plague challenge. *Infect. Immun.* 64:2180-2187.
19. **Anderson, G.W. Jr.**, S.E.C. Leary, E.D. Williamson, R.W. Titball, S.L. Welkos, P.L. Worsham, and A.M. Friedlander. 1996. Recombinant V antigen protects mice against pneumonic and bubonic plague caused by F1-capsule-positive and -negative strains of *Yersinia pestis*. *Infect. Immun.* 64:4580-4585.
20. **Anderson, G.W. Jr.**, P.L. Worsham, C.R. Bolt, G.P. Andrews, S.L. Welkos, A.M. Friedlander, and J.P. Burans. 1997. Protection of mice from fatal bubonic and pneumonic plague by passive immunization with monoclonal antibodies against the F1 protein of *Yersinia pestis*. *Am. J. Trop. Med. Hyg.* 64:4580-4585.
21. Heath, D.G., **G.W. Anderson, Jr.**, S.L. Welkos, A.M. Friedlander, and J.M. Mauro. 1997. A recombinant capsular F1-V antigen fusion protein vaccine protects against experimental bubonic and pneumonic plague. *in* *Vaccines 97*. Cold Spring Harbor Laboratory Press, pp 197-200.
22. Pullen, J.K., **G.W. Anderson, Jr.**, S.L. Welkos, and A.M. Friedlander. 1998. Analysis of the *Yersinia pestis* V protein for the presence of linear antibody epitopes. *Infect. Immun.* 66:521-527.

23. **Anderson, G.W. Jr.**, D.G. Heath, C.R. Bolt, S.L. Welkos, and A.M. Friedlander. Short- and long-term efficacy of single-dose subunit vaccines against *Yersinia pestis* in mice. *Am. J. Trop. Med. Hyg.*, 58(6): 793-799.
24. Ivins, B.E., M.L.M. Pitt, P.F. Fellows, J.W. Farchaus, G.E. Benner, D.M. Waag, S.F. Little, **G.W. Anderson, Jr.**, P.H. Gibbs, and A.M. Friedlander. 1998. Comparative efficacy of experimental anthrax vaccine candidates against inhalation anthrax in rhesus macaques. *Vaccine*, 16(11/12): 1141-1148.
25. Heath, D.G., **G.W. Anderson, Jr.**, J.M. Mauro, S.L. Welkos, G.P. Andrews, J. Adamovicz, and A.M. Friedlander. 1998. Protection against experimental bubonic and pneumonic plague by a recombinant capsular F1-V antigen fusion protein vaccine. *Vaccine*, 16(11/12): 1131-1137.
26. Andrews, G.P., S.T. Strachan, G.E. Benner, A.K. Sample, J.J. Adamovicz, **G.W. Anderson, Jr.**, S.L. Welkos, A.M. Friedlander. 1999. Protective efficacy of recombinant *Yersinia* outer proteins (Yops) against bubonic plague caused by encapsulated and non-encapsulated *Yersinia pestis*. *Infect. Immun.*, 67(3): 1533-1537.
27. Lawrence W.B., **G.W. Anderson, Jr.**, J.O. Lee, and W.C. Hall. Ocular sequelae associated with Rift Valley fever virus (RVFV) infection in inbred rats (submitted).

PUBLISHED ABSTRACTS:

1. Rosebrock, J.A., **G.W. Anderson, Jr.**, H. Schellekens, and C.J. Peters. 1983. Differential interferon sensitivities of lethal and non-lethal strains of Rift Valley fever virus (RVFV) in vitro. *In vitro* 19: 286-287.
2. **Anderson, G.W., Jr.**, M.V. Slayter, and C.J. Peters. 1988. Pathogenesis of a phleboviral infection (Punto Toro virus) in golden Syrian hamsters. *Virus Supplement* 2: 40.
3. Ribas, J.L., M.D. Kanzer, **G.W. Anderson, Jr.**, J. Sesterhenn, and C.J. Peters. 1989. Rift Valley fever viral encephalitis in the gerbil. *J. Neuropathol. Exp. Neurol.* 48: 315.
4. Ribas, J.L., M.D. Kanzer, **G.W. Anderson, Jr.**, E. Perez-Rosario, and C.J. Peters. 1990. Rift Valley fever viral encephalitis in the gerbil: Ultrastructural and immunocytochemical correlation. *J. Neuropath. Exp. Neurol.* 49:348(A).

Other publications:

Technical Report for Alternate Air Collection Media, SRS Technologies, TR98-156

PRESENTATIONS:

1. **Anderson, G.W., Jr.**, and J.V. Osterman. Susceptibility of mouse strains to *Rickettsia akari*. Presented at the 79th Annual Meeting of the American Society of Microbiologists, Los Angeles, California, 1979.
2. **Anderson, G.W., Jr.**, and J.V. Osterman. Susceptibility of mice to *Rickettsia akari*. Presented at the 1st Annual Rickettsiology Conference. Port Deposit, Maryland, 1979.
3. **Anderson, G.W., Jr.**, and C.J. Peters. Effect of immunosuppression on genetically resistant LEWIS/Mai rats to Rift Valley fever virus. Presented at the Maryland-D.C. Branch of the American Society for Microbiology Meeting, Fort Detrick, Frederick, Maryland, January 1981.
4. **Anderson, G.W., Jr.**, C.J. Peters, and T.W. Slone. Pathogenesis of Rift Valley fever virus in inbred rats. Presented at the 81st Annual Meeting of the American Society for Microbiology, Dallas, Texas, March 1981. Abstracts of the Meeting, D244.
5. Peters, C.J., and **G.W. Anderson, Jr.** Pathogenesis of Rift Valley fever and other Phlebovirus infections. Presented at the 5th International Congress of Virology, Strasbourg, France, August 1981.
6. Peters, C.J., and **G.W. Anderson, Jr.** Resistance to Phleboviruses. Presented at the U.S.-Japan Cooperative Medical Science Program, Bethesda, Maryland, 9 November 1981.
7. Peters, C.J., and **G.W. Anderson, Jr.** Pathogenesis of Phlebovirus infections. Presented at the 30th Annual Meeting of the American Society of Tropical Medicine and Hygiene, San Juan, Puerto Rico, November 1981.
8. **Anderson, G.W., Jr.**, and C.J. Peters. Role of humoral immunity in Rift Valley fever infection. Presented at the 49th Conjoint Meeting on Infectious Diseases, Ontario, Canada, 25 November 1981.
9. **Anderson, G.W., Jr.**, J.A. Rosebrock, A.J. Johnson, and C.J. Peters. Age- and dose-dependent resistance of rats to Rift Valley fever virus. Presented at the 82nd Annual Meeting of the American Society for Microbiology, Atlanta, Georgia, March 1982.
10. **Anderson, G.W., Jr.**, T.W. Slone, Jr., and C.J. Peters. A model for the encephalitic form of Rift Valley fever. Presented at the 31st Annual Meeting of the American Society of Tropical Medicine and Hygiene, Cleveland, Ohio, 1982.
11. Peters, C.J., H. Schellekens, J.A. Rosebrock, and **G.W. Anderson, Jr.** Genes, macrophages, and resistance to Rift Valley fever in the rat. Presented at the First Annual Meeting of the American Society for Virology, Ithaca, New York, 1982.

12. Peters, C.J., H. Schellenkens, J.A. Rosebrock, and **G.W. Anderson, Jr.** Genetic resistance to Rift Valley fever virus: Role of macrophages and interferon. Fourth International Conference on Comparative Virology, Banff, Canada, October 1982.
13. **Anderson, G.W., Jr.**, and J.F. Smith. Rift Valley fever virus (RVFV) maturation at the plasma membrane of rat hepatocytes as revealed by immunoelectron microscopy. Presented at the 35th Annual Meeting of the American Society of Tropical Medicine and Hygiene, Denver, Colorado; 8-11 December 1986.
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37. **Anderson, Jr. G.W.**, Laboratory/Field Biosafety. Presented at the Veterinary and Human Brucellosis Workshop, Almaty, Kazakhstan, 19-22 July 2004.
38. **Anderson, Jr. G.W.**, Collaborative Biosafety Efforts between the Defense Threat Reduction Agency (DTRA) and Russian Institutes. Presented at the Development of International Collaboration in Infectious Disease Research Conference, Novosibirsk, Russia, 8-10 September 2004.

File: F1-V fusion last update REDACTED
 Protocol: B94-02
 F1-V fusion protein immunization and challenge

File P 124-133

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath has produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Antigen dose will be based on 10 μ g F1/dose + the amount of V which is fused to it. Subcutaneous injection at nape of neck.

V-antigen used in this experiment is from Mauro. Details of the V-antigen can be obtained from CPT Heath's notebook # _____ page _____.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 strain of Y. pestis by sc and aerosol challenge, 50 LD₅₀.

EcF1c will be endotoxin free, same F1 as used in the active immunization experiment.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, _____ μ g of AL/dose

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

Groups 1-9, the amount of F1 will be held constant.

		Strain	#	Mice
Group 1	Alhydrogel alone, days 0, 30, sc	CO92	10	
Group 2	alhydrogel + 10 μ g F1, days 0, 30, sc	CO92	10	
Group 3	alhydrogel + 10 μ g F1 urea treated, days 0, 30, sc	CO92	10	
Group 4	Alhydrogel + 18.5 μ g F1-V fusion protein days 0, 30, sc	CO92	10	

Aerosol challenge

Group 5	Alhydrogel alone, days 0, 30, sc	CO92	10	
Group 6	alhydrogel + 10 μ g F1, days 0, 30, sc	CO92	10	
Group 7	alhydrogel + 10 μ g F1 urea treated, days 0, 30, sc	CO92	10	
Group 8	Alhydrogel + 18.5 μ g F1-V fusion protein, days 0, 30, sc	CO92	10	
Group 9	Alhydrogel + 37.0 μ g F1-V fusion protein days 0, 30, sc	CO92	10	
		Total	90	

1F622A6174/F-V-000

1F633B5D66/F-V-001

1F666A563B/F-V-002

1F684B1B13/F-V-003

1F73143D1D/F-V-004

1F6336192F/F-V-005

1F617A0402/F-V-006

1F6373612A/F-V-007

1F642A0E45/F-V-008

1F64667324/F-V-009

1F610F757C/F-V-010

1F625B356F/F-V-011

1F62584463/F-V-012

2007432B6B/F-V-013

1F620D6B07/F-V-014

1F65793A49/F-V-015

REDACTED

put

1F61041D5F/F-V-016
1F59570130/F-V-017
1F62776325/F-V-018
1F6E2E3015/F-V-019
1F60055D1F/F-V-020
1F66337F49/F-V-021
1F6E25735B/F-V-022
1F647B4141/F-V-023
1F6835251F/F-V-024
1F767F7A72/F-V-025, Not responding replaced with 1F617C1371
1F62530428/F-V-026
1F63717815/F-V-027
1F65122842/F-V-028
1F666F1F6D/F-V-029
1F56376371/F-V-030
1F66655A3C/F-V-031
1F663A6859/F-V-032
1F64163037/F-V-033
1F72107B64/F-V-034
1F63467B3D/F-V-035
1F645B1111/F-V-036
1F61524668/F-V-037
1F6132321C/F-V-038
1F655A0E14/F-V-039
1F661E3726/F-V-040, Not responding, replaced with 1F6134004C
1F656C5937/F-V-041
1F61135D10/F-V-042
1F6655574F/F-V-043
1F664B210F/F-V-044
7F7B0A2C5B/F-V-045
1F63737516/F-V-046
1F66240C4B/F-V-047
1F5A6F0C0C/F-V-048
1F65354106/F-V-049
1F76011A50/F-V-050
1F653D7946/F-V-051
1F624D6A48/F-V-052
1F64030B6F/F-V-053
1F6E295D6D/F-V-054
1F64303B12/F-V-055
1F635D326F/F-V-056
1F6726084C/F-V-057
1F6317796E/F-V-058
1F64024437/F-V-059
1F66056C0A/F-V-060, died from anesthesia on 17FEB95 during bleeding
1F637E6917/F-V-061
1F64653F59/F-V-062
1F643F7846/F-V-063
1F6412204B/F-V-064
7F7D261230/F-V-065
1F6132735B/F-V-066
1F635C5D45/F-V-067
1F636A1103/F-V-068
1F5F542F7F/F-V-069

1F6729527F/F-V-070
 1F63280353/F-V-071
 1F600B6D09/F-V-072
 1F77037275/F-V-073
 1F615E071B/F-V-074
 1F646F0D01/F-V-075
 1F66415A60/F-V-076
 1F65566046/F-V-077
 1F64495262/F-V-078
1F6467682E/F-V-079
 200905054D/F-V-080
 1F636F2F60/F-V-081
 1F650E541A/F-V-082
 1F637B3E45/F-V-083
 1F673C5668/F-V-084
 1F647E2E51/F-V-085
 1F663B0937/F-V-086
 1F62111658/F-V-087
 1F6320124C/F-V-088
 1F635A1F05/F-V-089

CFA from Sigma Cat#F-5881 Lot#80H8808, 10 ml/bottle

IFA from Sigma Cat#F-5506 Lot#80H8812, 10 ml/bottle

Groups 10-23 will be started one week later. The amount of V will be held constant, though the V part of the fusion protein is only about half the size of the native V.

Subcutaneous challenge

		Strain	# Mice
Group 10	Alhydrogel alone, days 0, 30, sc	C12	10
Group 11	Alhydrogel days + 27 µg F1-V fusion protein day 0, 30, sc	C12	10
Group 12	CFA + 10 µg V, days 0, IFA 30, ip	C12	10
Group 13	CFA + 10 µg V urea treated, days 0, IFA 30, ip	C12	10
Group 14	CFA + 27 µg F1-V fusion protein days 0, IFA 30, ip	C12	10
Group 15	CFA + 54 µg F1-V fusion protein days 0, IFA 30, ip	C12	10
Group 16	CFA day 0, IFA day 30 alone, ip	C12	10

Aerosol challenge

Group 17	Alhydrogel alone, days 0, 30, sc	C12	10
Group 18	Alhydrogel + 27 µg F1-V fusion protein, days 0, 30, sc	C12	10
Group 19	CFA + 10 µg V, days 0, IFA 30, ip	C12	10
Group 20	CFA + 10 µg V urea treated, days 0, IFA30, ip	C12	10
Group 21	CFA + 27 µg F1-V fusion protein day 0, IFA 30, ip	C12	10
Group 22	CFA + 54 µg F1-V fusion protein day 0, IFA 30, ip	C12	10
Group 23	CFA day 0, IFA day 30 alone, ip	C12	10
		Total	140

Group 24 CFA + 27 µg F1-V fusion protein day 0, IFA 30, ip --antibody response 10
 Measure titer at 14, 27,57

Chip # for Groups 10-24, Group 10 has some mice doubled

1F646A0D06/F-V-090

7F7B107C58/F-V-091 or 1F63125319, DOUBLE CHIPPED

1F6E345D62/F-V-092 OR 1F64791E66, DOUBLE CHIPPED

1F64141752/F-V-093 OR 1F61116E01, DOUBLE CHIPPED
1F65020D6D/F-V-094 OR 9F7D25797E, DOUBLE CHIPPED
1F62032B51/F-V-095 OR 1F650627AF, DOUBLE CHIPPED
1F635D4B56/F-V-096 OR 7F7D243700, DOUBLE CHIPPED
7F7B06623C/F-V-097 OR 7F7D17224D, DOUBLE CHIPPED
7F7D23252D/F-V-098
1F630A7004/F-V-099
1F6458061F/F1-VB-001
1F66294012/F1-VB-002
1F65323119/F1-VB-003
1F68597E22/F1-VB-004
1F663B023E/F1-VB-005
1F64742F5A/F1-VB-006
1F62713757/F1-VB-007
1F664A1B16/F1-VB-008
1F624C3D76/F1-VB-009
1F60466754/F1-VB-010
1F650D343B/F1-VB-011
1F640F0668/F1-VB-012
1F684A6D42/F1-VB-013
1F627B4440/F1-VB-014
1F66362421/F1-VB-015
1F64121C4F/F1-VB-016
1F647A4340/F1-VB-017
1F757C7977/F1-VB-018
1F65625644/F1-VB-019
1F65662E68/F1-VB-020
1F655A455D/F1-VB-021, found dead, 13JAN95, cause unknown
1F63624B51/F1-VB-022
1F65487440/F1-VB-023
1F61185F09/F1-VB-024
1F63296273/F1-VB-025
1F6626094C/F1-VB-026
1F622A1C39/F1-VB-027
1F6122312D/F1-VB-028
1F6329775E/F1-VB-029
1F64652276/F1-VB-030
1F73061751/F1-VB-031
1F63672C6B/F1-VB-032
1F68406158/F1-VB-033
1F6359061F/F1-VB-034
1F653E2C12/F1-VB-035
1F60524B64/F1-VB-036
1F67274A09/F1-VB-037
200F13231B/F1-VB-038
1F620B7B79/F1-VB-039
1F66121653/F1-VB-040
1F66247661/F1-VB-041
1F652F0D40/F1-VB-042
1F63103B33/F1-VB-043
1F673D0C31/F1-VB-044
1F62757218/F1-VB-045
1F63401727/F1-VB-046
1F665F5D3F/F1-VB-047

1F66585F44/F1-VB-048
1F61195611/F1-VB-049
200737366C/F1-VB-050
1F614E2012/F1-VB-051
1F64297C58/F1-VB-052
1F75463175/F1-VB-053
1F626E157C/F1-VB-054
1F650F5419/F1-VB-055
1F64075C1A/F1-VB-056
1F756A3151/F1-VB-057
1F6121124D/F1-VB-058
1F655E405E/F1-VB-059
1F65233C1D/F1-VB-060
1F683C0B32/F1-VB-061
1F615F021F/F1-VB-062
1F71772257/F1-VB-063
1F644F713D/F1-VB-064
1F67412415/F1-VB-065
1F66606C2F/F1-VB-066
1F77087969/F1-VB-067
1F637D1F62/F1-VB-068
1F61172742/F1-VB-069
1F64511E0E/F1-VB-070
1F5756496B/F1-VB-071
1F681C4617/F1-VB-072
1F68571012/F1-VB-073
1F682F6763/F1-VB-074
1F673E4577/F1-VB-075
1F62681205/F1-VB-076
1F655F1E7F/F1-VB-077
1F65523377/F1-VB-078
1F61284810/F1-VB-079
1F6152307E/F1-VB-080
1F68382A17/F1-VB-081
1F686B1876/F1-VB-082
1F60106E03/F1-VB-083
1F63242D2D/F1-VB-084
1F637A7311/F1-VB-085
1F631F0D52/F1-VB-086
1F6378473F/F1-VB-087
1F63390C39/F1-VB-088
1F66517931/F1-VB-089
1F642C272A/F1-VB-090
1F654E703E/F1-VB-091
1F64134129/F1-VB-092
1F63355772/F1-VB-093
1F621A0263/F1-VB-094
1F64361A2D/F1-VB-095
1F61774A3F/F1-VB-096
1F65276075/F1-VB-097
1F646F0905/F1-VB-098
1F624C181B/F1-VB-099
1F684E3C6F/F1-VB-100
1F64034B2F/F1-VB-101

1F65083C38/F1-VB-102
1F65792E55/F1-VB-103
1F6012412E/F1-VB-104
1F68321F28/F1-VB-105
1F6E313909/F1-VB-106
1F614D6F44/F1-VB-107
1F62394402/F1-VB-108
1F627B5C28/F1-VB-109
1F64165A0D/F1-VB-110
1F630B4231/F1-VB-111
1F6629450D/F1-VB-112
1F5F630D12/F1-VB-113
1F657C5B25/F1-VB-114
1F6337596E/F1-VB-115
1F66297260/F1-VB-116
1F66524F5A/F1-VB-117
1F6829537D/F1-VB-118
1F64616E2E/F1-VB-119
1F61517936/F1-VB-120
1F65136504/F1-VB-121
1F66002A51/F1-VB-122
1F5F7D1075/F1-VB-123
1F65074134/F1-VB-124
1F71724836/F1-VB-125
1F656C4F41/F1-VB-126
1F60182148/F1-VB-127
1F6866573C/F1-VB-128
1F62780205/F1-VB-129
1F65080173/F1-VB-130

Schedule

Groups 1-9

22NOV94 Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in AA-3 (Barrier)
01DEC94 Chipped with BioMedic Data Systems transponders in AA-3
16DEC94 1st immunization, AA-3
13JAN95 2nd Immunization, AA-3, day 28
17FEB95 Bleed to determine prechallenge titers, AA-3, day 63, serum#1500-1589
24FEB95 Challenge by aerosol & sc routes, day 70
24MAR95 Terminal bleed, day 28 pi, titrate spleens, serum # 2403-2453
Term # 9378 - 9432

Groups 10-23

29NOV94 Arrival of Swiss Webster mice, female 7-8 wks, Harland Sprague Dawley in AA-3 (Barrier)
13DEC94 Chipped with BioMedic Data Systems transponders in AA-3 (SGT Zimmerman, Vet Med)
22DEC94 1st immunization, AA-3; Crow & Fitzgerald helped
20JAN95 2nd immunization, AA-3, day 29
24FEB95 Bleed to determine prechallenge titers, AA-3, day 64, serum# 1628-1708
3MAR95 Challenge by aerosol & sc route, day 71
31MAR95 Terminal bleed, day 28 pi, titrate spleens, serum # 1759-2800
Term # 9433-9479

Group 24

22DEC94 1st immunization, AA-3
06JAN95 Bleed, day 14, AA-3, SERUM# 1220-1229
20JAN95 Bleed, day 27, AA-3, SERUM# 1339-1348
20JAN95 2nd immunization, day 28, AA-3
24FEB95 Bleed, day 63, AA-3, SERUM# 1768-1778 and bronchial lavage #

see page 123

REDACTED	Project: Active Immunization F1-V Alhydrogel																										
Notebook #: 3598																											
Inoculum: <i>Yersinia pestis</i> strain CO92																											
Route: aerosol Dose: RUN 1 = 80 LD ₅₀ RUN 2 = 104 LD ₅₀																											
REDACTED	Swiss Webster																										
	Day	Age:	REDACTED	at 7-wk	Vendor: Harlan	Sprague Dawley	Sex: female																				
				7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25					
Day postinfection	0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	
Group	Cage#																										
Alh alone	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
GP5	2	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	3	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	4	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	5	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	6	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	7	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	8	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	9	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	10	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Alh+F1	11	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
10 µg	12	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
GP6	13	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	14	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	15	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	16	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	17	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	18	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	19	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	20	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Alh+F1	21	Died during bleeding of 17FEB95																									
10 µg	22	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
urea	23	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
GP7	24	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	25	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	26	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	27	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	28	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	29	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	30	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Alh+F1-V	31	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
18.5 µg	32	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
GP8	33	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	34	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	35	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	36	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	37	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	38	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	39	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	40	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
Alh+F1-V	41	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
37 µg	42	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
GP9	43	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	44	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	45	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	46	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	47	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	48	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	49	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
	50	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
REDACTED		Comments/Chip #																									
GREEN		BLUE																									
RED		BLACK																									

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Use scanner to check chip # of dead animals

Use scanner to check chip
Mark number of mice alive

Attn: LTC Anderson

Date REDACTED	Project: Active Immunization F1-V Alhydrogel/CFA			
Notebook #: 3598				
Inoculum: Yersinia pestis strain C12				
Route: SC	Dose:	REDACTED		
REDACTED	Miss Webster	Age: REDACTED	at 7-8wk	Vendor: Harlan Sprague Dawley
	3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 1			Sex: female
Day postinfection	0 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29			Comments/Chip #
Group	Cage#			
Alh alone	GP10			1F646A0D06/F-V-090 7F7B107C58/F-V-091 or 1F6 1F6E345D62/F-V-092 OR 1F6 1F64141752/F-V-093 OR 1F6 1F65020D6D/F-V-094 OR 9F 1F62032B51/F-V-095 OR 1F6 1F635D4B56/F-V-096 OR 7F 7F7B06623C/F-V-097 OR 7F 7F7D23252D/F-V-098 1F630A7004/F-V-099
Alh+F1-V 27 µg	GP11			1F6458061F/F1-VB-001 1F66294012/F1-VB-002 1F65323119/F1-VB-003 1F68597E22/F1-VB-004 1F663B023E/F1-VB-005 1F64742F5A/F1-VB-006 1F62713757/F1-VB-007 1F664A1B16/F1-VB-008 1F624C3D76/F1-VB-009 1F60466754/F1-VB-010
CFA+V 10 µg	GP12			1F650D343B/F1-VB-011 1F640F0668/F1-VB-012 1F684A6D42/F1-VB-013 1F627B4440/F1-VB-014 1F66362421/F1-VB-015 1F64121C4F/F1-VB-016 1F647A4340/F1-VB-017 1F757C7977/F1-VB-018 1F65625644/F1-VB-019 1F65662E68/F1-VB-020
CFA+V 10 µg urea	GP13	Found dead, 13JAN95, cause unknown		1F655A455D/F1-VB-021 1F63624B51/F1-VB-022 1F65487440/F1-VB-023 1F61185F09/F1-VB-024 1F63296273/F1-VB-025 1F6626094C/F1-VB-026 1F622A1C39/F1-VB-027 1F6122312D/F1-VB-028 1F6329775E/F1-VB-029 1F64652276/F1-VB-030
CFA-F1-V 27 µg	GP14			1F73061751/F1-VB-031 1F63672C6B/F1-VB-032 1F68406158/F1-VB-033 1F6359061F/F1-VB-034 1F653E2C12/F1-VB-035 1F60524B64/F1-VB-036 1F67274A09/F1-VB-037 200F13231B/F1-VB-038 1F620B7B79/F1-VB-039 1F66121653/F1-VB-040
CFA-F1-V 54 µg	GP15			1F66247661/F1-VB-041 1F652F0D40/F1-VB-042 1F63103B33/F1-VB-043 1F673D0C31/F1-VB-044 1F62757218/F1-VB-045 1F63401727/F1-VB-046 1F665F5D3F/F1-VB-047 1F66585F44/F1-VB-048 1F61195611/F1-VB-049 200737366C/F1-VB-050

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Discard dead animals

Use scanner to check chip # of dead animals

Mark number of mice alive

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Use scanner to check chip # of dead animals

Mark number of mice alive

ELISA, F1 & VANT

STARTING DILUTION 1:640, SERIAL 1:2 DILUTIONS

F1 AND V ANTIGENS (

<u>F1 PLATE #</u>	<u>SERUM #</u>	<u>V PLATE #</u>	<u>F1 PLATE #</u>	<u>SERUM #</u>	<u>V PLATE #</u>
1A F	1117	16A	11A	1844	26A
B	1118	16B	B	1845	B
2A	1119	17A	12A	1846	27A
B	1230	17B	B	1847	B
3A	1231	18A	13A	1848	28A
B	1232	18B	B	1849	B
4A	1233	19A	14A	1850	29A
B	1234	19B	B	NEG(N.M.)	B
5A	1235	20A	15A	F1+	30A
B	1236	1836(F1)	20B	B	831
6A	1237	1837(F1)	21A		
B	1238	1838(F1)	21B		
7A	1836	1236(F1)	22A	PLATE 1-3(F1):	
B	1837	1237(F1)	22B	200 μ l 10 10	
8A	1838	1238(F1)	23A	100 110 110	
B	1839	1839	23B	REMOVED BUFFER FROM COLUMNS 2-12	
9A	1840	1840	24A	ADDED 100 μ l EACH SAMPLE TO COLUMN 1, 100 μ l BUFFER	
B	1841	1841	24B	TO COLUMNS 2-12, & RETITRATED SAMPLES.	
10A	1842	1842	25A		
B	1843	1843	25B		

SUMMARY

			F1	V
1836	POOL GP10	Alhydrogel alone	Day63 active F1-V	0
1837	POOL GP11	Alh+27ugF1-V		81820
1838	POOL GP12	CFA+10ugV	Day63 active F1-V	0
1839	POOL GP13	CFA+10ugV,urea	Day63 active F1-V	1310720
1840	POOL GP14	CFA+27ugF1-V	Day63 active F1-V	655360
1841	POOL GP15	CFA+54ugF1-V	Day63 active F1-V	163840
1842	POOL GP16	CFA alone	Day63 active F1-V	1310720
1843	POOL GP17	Alhydrogel alone	Day63 active F1-V	1280
1844	POOL GP18	Alh+27ugF1-V	Day63 active F1-V	2560
1845	POOL GP19	CFA+10ugV	Day63 active F1-V	40960
1846	POOL GP20	CFA+10ugV,urea	Day63 active F1-V	81820
1847	POOL GP21	CFA+27ugF1-V	Day63 active F1-V	655360
1848	POOL GP22	CFA+54ugF1-V	Day63 active F1-V	163840
				327680
				327680

REDACTED

• Data on page 131 in the first section evidence that the F1-V fusion protein can induce an immune response to both the F1 and V portion of the F1-V fusion protein. This is the first proof of the concept of making a fusion protein which could be used as an immunogen in a future plague vaccine. There was some non-specific reactivity toward V, but low levels compared to the F1-V treated animals.

The second part of the proof will be the protection studies started on [REDACTED] and [REDACTED] of the protection studies with the C092 challenge already shown protection, then other fusion proteins which are unique could significantly reduce the complexity of the manufacturing process. F1-V forming an aggregate, may also increase the immunogenicity of V.

[REDACTED]

David M. Heath
Donald L. Johnson

Summary [REDACTED]

Serum#	Group	Treatment	Bleed Date	Day Post	F1 Titer		V Titer		Change(wells)
1836	POOL GP10	Alhydrogel alone	[REDACTED] ED	Day63	0	0	0	0	0
1837	POOL GP11	Alh+27ugF1-V		Day63	40960	81920	163840	1	
1838	POOL GP12	CFA+10ugV		Day63	0	1310720	1310720	0	
1839	POOL GP13	CFA+10ugV,urea		Day63	0	655360	1310720	1	
1840	POOL GP14	CFA+27ugF1-V		Day63	163840	1310720	1310720	0	
1841	POOL GP15	CFA+54ugF1-V		Day63	163840	1310720	1310720	0	
1842	POOL GP16	CFA alone		Day63	0	1280	1280	0	
1843	POOL GP17	Alhydrogel alone		Day63	0	2560	0	-2	
1844	POOL GP18	Alh+27ugF1-V		Day63	40960	81920	163840	1	
1845	POOL GP19	CFA+10ugV		Day63	0	655360	1310720	1	
1846	POOL GP20	CFA+10ugV,urea		Day63	0	1310720	1310720	0	
1847	POOL GP21	CFA+27ugF1-V		Day63	163840	655360	1310720	1	
1848	POOL GP22	CFA+54ugF1-V		Day63	327680	327680	1310720	2	
1849	POOL GP23	CFA alone		Day63	0	10240	1280	-3	
1850	POOL GP24	CFA+27ugF1-V		Day63	163840	163840	1310720	3	
Controls:									
		F1+ Pool		40960		20480	10240	-1	
		831		0		163840	655360	2	
		Normal Mouse		0		5120	0	-4	

new page 131 2/12/3

file - Pitt-aerosol data. [REDACTED]

EXPT. #3: [REDACT] Aerosol and sc Plague Challenge Expt. (mice)

FOR ACTIVE F1-V IMMUNIZATION C14111EN C12 3 MAR 95
AEROSOL - SITE PAGES 127 - 130

	Target		Calculated	
suspension	Conc./ml	no. CFU/ml	inhaled no. CFU	No. LD50s
Prespray	1.75x10e10/ml	2.8 x 10e10/ml		
C092/C12				

AGI
1
2
3

2.5 x 10e8/ml
2.6 x 10e8/ml
3.4 x 10e8/ml

new page 133
N 1050 plaque

SUBCUTANEOUS -

Target

Date: **REDACTED**PI: LTC Anderson
Agent: Plague
Strain: C12

Animal Model: Mouse

C12 LD50=1.1E+05

Wt: (Ave.): 27.69

Sex: female

				Inhaled Dose		
AGI/ml	AGI	aerosol	MV	cfu	LD50s	Strain
2.50E+08	2.50E+09	4.17E+07	0.025	1.04E+07	94.70	C12
2.60E+08	2.60E+09	4.33E+07	0.025	1.08E+07	98.48	C12
3.40E+08	3.40E+09	5.67E+07	0.025	1.42E+07	128.79	C12

Summary **REDACTED**

Mouse weight

27.0

32.2

23.4

28.1

25.6

25.6

29.0

30.5

30.1

35.4

37.69g avg

#	Group	Treatment	Blood date	File: Serumbook	File Update: REDACTED	F1 TITER	V TITER
				Bleed day	Protocol		
220	GP24	F1-V, day 14 antibody	REDACTED	DAY14	F1-V ANTIBODY ONTOGENY	81920	40960
221	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	20480	40960
222	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	81920	20480
223	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	20480	10240
224	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	40960	640
225	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	10240	0
226	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	40960	10240
227	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	40960	40960
228	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	20480	10240
229	GP24	F1-V, day 14 antibody		DAY14	F1-V ANTIBODY ONTOGENY	20480	20480
Geomean(of positive values only!)						31042	13934

340	GP24 F1-V, day 14 antibody	REDACTED	DAY28	F1-V ANTIBODY ONTOGENY	163840	327680	
			Bleed day	Protocol	F1 TITER	V TITER	
341	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	81920	327680	
342	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	81920	1280	
343	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	81920	1310720	
344	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	20480	2560	
345	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	81920	327680	
346	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	327680	655360	
347	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	81920	40960	
348	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	40960	81920	
349	GP24 F1-V, day 14 antibody		DAY28	F1-V ANTIBODY ONTOGENY	81920	163840	
Geomean(of positive values only!)						81920	94101

768	GP24 CFA+27ugF1-V	REDACTED	Day63	active F1-V	327680	1310720
			Bleed day	Protocol	F1 TITER	V TITER
769	GP24 CFA+27ugF1-V		Day63	active F1-V	655360	655360
770	GP24 CFA+27ugF1-V		Day63	active F1-V	327680	655360
771	GP24 CFA+27ugF1-V		Day63	active F1-V	327680	1310720
772	GP24 CFA+27ugF1-V		Day63	active F1-V	655360	655360
773	GP24 CFA+27ugF1-V		Day63	active F1-V	327680	1310720
774	GP24 CFA+27ugF1-V		Day63	active F1-V	163840	40960
775	GP24 CFA+27ugF1-V		Day63	active F1-V	163840	1310720
776	GP24 CFA+27ugF1-V		Day63	active F1-V	655360	1310720
777	GP24 CFA+27ugF1-V		Day63	active F1-V	Died	

Geomean(of positive values only!) 353914 707828

Summary REDACTED

File: Serumbook				File Update:	REDACTED			
Serum #	Group	Treatment	Bleed date	Bleed day	Protocol	CHIP #	MISC	V TITER
1688	GP16	CFA alone	REDACTED	Day63	active F1-V	1F614E2012/F1-VB-051		O
1689	GP16	CFA alone		Day63	active F1-V	1F64297C58/F1-VB-052		O
1690	GP16	CFA alone		Day63	active F1-V	1F75463175/F1-VB-053		O
1691	GP16	CFA alone		Day63	active F1-V	1F626E157C/F1-VB-054		O
1692	GP16	CFA alone		Day63	active F1-V	1F650F5419/F1-VB-055		O
1693	GP16	CFA alone		Day63	active F1-V	1F64075C1A/F1-VB-056		O
1694	GP16	CFA alone		Day63	active F1-V	1F756A3151/F1-VB-057	No sample	
1695	GP16	CFA alone		Day63	active F1-V	1F6121124D/F1-VB-058		640
1696	GP16	CFA alone		Day63	active F1-V	1F655E405E/F1-VB-059		O
1697	GP16	CFA alone		Day63	active F1-V	1F65233C1D/F1-VB-060		O
Geomean(of positive values only!)								640

1758	GP23	CFA alone	REDACTED	Day63	active F1-V	1F65136504/F1-VB-121		1280
1759	GP23	CFA alone		Day63	active F1-V	1F66002A51/F1-VB-122		O
1760	GP23	CFA alone		Day63	active F1-V	1F5F7D1075/F1-VB-123	20480	
1761	GP23	CFA alone		Day63	active F1-V	1F65074134/F1-VB-124		O
1762	GP23	CFA alone		Day63	active F1-V	1F71724836/F1-VB-125		640
1763	GP23	CFA alone		Day63	active F1-V	1F656C4F41/F1-VB-126		O
1764	GP23	CFA alone		Day63	active F1-V	1F60182148/F1-VB-127		O
1765	GP23	CFA alone		Day63	active F1-V	1F6866573C/F1-VB-128		O
1766	GP23	CFA alone		Day63	active F1-V	1F62780205/F1-VB-129	1280	
1767	GP23	CFA alone		Day63	active F1-V	1F65080173/F1-VB-130		O
Geomean(of positive values only!)								2153

Controls:

Normal Mouse	O
F1+ Pool	5120
Serum 831	327680

CFA appear to exceed background pattern. # 1760 is so high, it appear to be an error of some sort. Most probable, a tube contaminated or mislabeled later.

Summary REDACTED

File: Serumbook				File Update:	REDACTED		
Serum #	Group	Treatment	Bleed date	Bleed day	Protocol	F1 TITER	V TITER
2081	GP1 Pool	Alhydrogel only	REDACTED	Day63	ALH-F1-V fusion	O	O
2082	GP2 Pool	ALH+10 μ g F1		Day63	ALH-F1-V fusion	81920	O
2083	GP3 Pool	ALH+10 μ g F2urea		Day63	ALH-F1-V fusion	81920	O
2084	GP4 Pool	ALH+18.5 μ g F1-V		Day63	ALH-F1-V fusion	81920	163840
2085	GP5 Pool	Alhydrogel only		Day63	ALH-F1-V fusion	O	O
2086	GP6 Pool	ALH+10 μ g F1		Day63	ALH-F1-V fusion	40960	O
2087	GP7 Pool	ALH+10 μ g F2urea		Day63	ALH-F1-V fusion	81920	O
2088	GP8 Pool	ALH+18.5 μ g F1-V		Day63	ALH-F1-V fusion	81920	327680
2089	GP9 Pool	ALH+37 μ g F1-V		Day63	ALH-F1-V fusion	163840	163840

AEROSOL EXPOSURE SHEET

gross exposure #: 95-0-9 H

Date: REDACTED

Agent Plague

Protocol

P.L. Anderson

see page 127-128

Wood Operator: S6, T David A. McCloskey

Brand Operator: SGT David A. McCloskey
Date of Birth: 08/01 DOB: 8/1 Wet T: 74 Rel. Hum.: 72%

ground system: No \rightarrow Only

Flow Rate: 12 L/min

Collision #: B

AEROSOL EXPOSURE SHEET

Aerosol exposure #: 95-032 H

Date: **REDACTED**

Aerosol Operator: BT

Pre-operational Check Performed: ✓ Dry T: 82 Wet T: 72 Rel. Hum.: 64%

Agent: PLAGUE

Aerosol System: NOSE - ONLY

Protocol F:

System Flow Rate: 12 LPM

P.L: LTC ANDERSON'

Collision #: B

Exhibit GA5

Dear Dr Friedlander

CBDE/USAMRIID COLLABORATIVE RESEARCH INTO PROTECTIVE EFFICACY OF RECOMBINANT V-ANTIGEN AGAINST PARENTERAL AND AEROSOL CHALLENGE WITH YERSINIA PESTIS

As you are aware, CBDE has data to suggest that the V-antigen of the plague causing organism *Yersinia pestis*, when used as an immunogen, is highly protective against plague. The V-antigen could therefore be a major component of an improved plague vaccine to be developed in the future by CBDE.

You recently indicated to us that USAMRIID wished to collaborate in testing the protective capacity of the V-antigen against parenteral and aerosol challenge with virulent plague. We agreed that such a collaboration would be desirable because it could generate valuable data which would be of benefit to both CBDE and USAMRIID. We therefore decided that the collaboration should, in the future, be the subject of a Project Arrangement under the Memorandum of Understanding between the Secretary of Defense (US) and the Secretary of State for Defence (UK) concerning Technology Research and Development Projects (which is currently still under negotiation).

However, we also agreed that any delay in the collaboration would reduce the benefit of the resulting data, and therefore it would be desirable to commence work in advance of a more formal Project Arrangement.

Accordingly, this letter sets out below the respective duties, rights and responsibilities of each of us under the collaboration, *pro tem*, pending the negotiation of a more comprehensive arrangement:

1. SCOPE OF WORK

a. CBDE will supply to USAMRIID, for the purposes described in (b), the following:

- i. 30 mg of recombinant *Yersinia pestis* V-antigen.
- ii. Protocols detailing the immunisation route, doses and schedules used at CBDE.
- iii. Polyclonal antisera raised against the V antigen of *Yersinia pestis*.
- iv. Details of the CBDE challenge route, challenge strain and protection data afforded by the V-antigen vaccine against parenteral challenge with *Yersinia pestis*.

b. USAMRIID will:

- i. Immunise groups of animals parenterally with the following:
 - V-antigen in combination with Alhydrogel.

Exhibit GA6

Suggested protocol for the F1-whole V fusion protein.

File: F1-wholeV fusion last update **REDACTED**

Protocol: B95-01

F1-WV fusion protein immunization and challenge

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Only part of the V-protein was used in the initial F1-V fusion. This F1-V fusion was immunogenic, but was not very efficacious when compared to the whole V-protein (WV) produced by Mauro to protect against F1 minus strains of *Y. pestis*. This is a repeat of part of the initial F1-V protection study using the whole V-protein in the F1-WV fusion. Subcutaneous injection along back of the mouse for immunization.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 and C12 strain of *Y. pestis* by sc and aerosol challenge, 100-Max LD₅₀.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, _____ μg of AL/dose

Endotoxin level in the F1-WV preparation is _____ U/ml.

Will use Mauro's V which has been urea treated as per the F1-WV procedure. Details in CPT Heath's laboratory notebook.

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

Implantable Micro Identification transponders from: BioMeric Data Systems, Inc 255 W. Spring Valley Ave. Maywood, NJ 07607, 1-800-526-BMDS

		Dose		
		Strain	LD ₅₀	# Mice
Subcutaneous challenge				
Group 1	Alhydrogel alone, days 0, 30, sc	C12	100	10
Group 2	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	C12	100	10 X
Group 3	Alhydrogel + 10 μg Mauro-V urea, days 0, 30, sc	C12	Max	10 X
Group 4	Alhydrogel + 13.6 μg F1-WV fusion protein days 0,30,sc	C12	Max	10 X
Group 5	Alhydrogel + 27.2 μg F1-WV fusion protein days 0, 30, sc	C12	Max	10 X
Group 6	Alhydrogel alone days 0, 30, sc	C12	Max	10 X
Group 7	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	CO92	100	10 X
Group 8	Alhydrogel alone, days 0, 30, sc	CO92	100	10

Aerosol challenge

Group 09	Alhydrogel alone, days 0, 30, sc	C12	50	10
Group 10	Alhydrogel + 13.6 μg F1-WV fusion protein day 0, 30, sc	C12	50	10 X
Group 11	Alhydrogel + 10 μg Mauro-V urea, days 0, 30, sc	C12	Max	10 X
Group 12	Alhydrogel + 13.6 μg F1-WV fusion protein, days 0, 30, sc	C12	Max	10 X
Group 13	Alhydrogel + 27.2 μg F1-WV fusion protein days 0,30, sc	C12	Max	10 X
Group 14	Alhydrogel alone, days 0, 30, sc	C12	Max	10 X
Group 15	Alhydrogel + 13.6 μg F1-WV fusion protein days 0, 30, sc	CO92	100	10 X
Group 16	Alhydrogel alone days 0, 30, sc	CO92	100	10 X
Group 17	Alhydrogel + 13.6 μg F1-WV fast prep, days 0, 30, sc	C12	Max	10 X
Group 18	Alhydrogel + 10 μg F1 + 10 μg Mauro's V, days 0, 30, sc	C12	Max	10 X
Group 19	Greer plague vaccine, days 0, 30, sc	C12	Max	10 05
Group 20	Alhydrogel alone, day 0, 30, sc	C12	Max	05

100

10	Group 21	ALH + 13.6 µg F1-WV fusion protein day 0, 30, sc --antibody response	10
		Measure titer at 7, 14, 27, 57, 90	
10	Group 22	ALH + 13.6 µg F1-WV fusion protein day 0, 30, sc --lung lavage, day 57	05
10	Group 23	ALH + 27.2 µg F1-WV fusion protein day 0, 30, sc --antibody response	10
		Measure titer at 7, 14, 27, 57, 90	
10	Group 24	ALH + 27.2 µg F1-WV fusion protein day 0, 30, sc --lung lavage, day 57	05
		ALH + Mauro-V urea, 10 ug, day 0, 30, sc --antibody response	
	Group 25	ALH + Mauro-V urea, 10 ug, day 0, 30, sc --lung lavage, day 57	05
	Group 26	ALH alone, day 0, 30, sc	10
	Group 27	Measure titer at 7, 14, 27, 57, 90	
10	Group 28	ALH alone, day 0, 30, sc, lung lavage, day 57	05
	Group 29	ALH alone, for spleen weights 28 day pi	10 05
			Total 220

Schedule

Groups 1-20

13Jun95 Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in AA-3 (Barrier)
 24Jun95 Chipped with BioMedic Data Systems transponders, West
 27Jun95 1st immunization, day 0
 27Jul95 2nd Immunization, day 30
 24Aug95 Bleed to determine prechallenge titers, day 58
 31Aug95 Challenge by aerosol & sc routes, day 65
 28Sep95 Terminal bleed, day 28 pi, titrate spleens# serum #

Group 21-25

13Jun95 Mice arrive
 27Jun95 1st immunization, AA-3
 11Jul95 Groups 21, 23, 25; Bleed, day 14, AA-3, SERUM#
 26Jul95 Groups 21, 23, 25; Bleed, day 29, AA-3, SERUM#
 27Jul95 2nd immunization, day
 31Aug95 Bleed, day 65, AA-3, Groups SERUM#
 Groups 22, 24, 26, and 28 for serum titer & bronchial lavage #
 28Sep95 Group 29 for Spleen weights #
 25Sep95 Groups 21, 23, and 25; day 90, serum #

Chip numbers for all groups extra alhydrogel controls

File: F1-wholeV fusion last update **REDACTED**
 Protocol: B95-01
 F1-WV fusion protein immunization and challenge

Investigators: CPT Heath, Dr. Welkos, LTC Anderson, COL Friedlander

Background. CPT David Heath produced and purified a recombinant F1-V fusion protein. The protein is positive by Western blot to F1 and V. Only part of the V-protein was used in the initial F1-V fusion. This F1-V fusion was immunogenic, but was not very efficacious when compared to the whole V-protein (WV) produced by Mauro to protect against F1 minus strains of *Y. pestis*. This is a repeat of part of the initial F1-V protection study using the whole V-protein in the F1-WV fusion. Subcutaneous injection along back of the mouse for immunization.

Purpose: Immunize and challenge mice to check on immunogenicity and protection against the CO92 and C12 strain of *Y. pestis* by sc and aerosol challenge, 100-Max LD₅₀.

Alhydrogel, 1.3%, from SuperFos. Batch # 2043, Expiration date None, 0.1857 mg of AL/dose

Endotoxin level in the F1-WV preparation is _____ U/ml.

Will use Mauro's V which has been urea treated as per the F1-WV procedure. Details in CPT Heath's laboratory notebook.

Immunization Groups: 10 Swiss Webster female mice per group from Harlan Sprague Dawley

Implantable Micro Identification transponders from: BioMeic Data Systems, Inc 255 W. Spring Valley Ave. Maywood, NJ 07607, 1-800-526-BMDS

				Dose	
			Strain	LD ₅₀	# Mice
Subcutaneous challenge					
Group 1	Alhydrogel alone, days 0, 30, sc		C12	100	10 NC
Group 2	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc		C12	100	10
Group 3	Alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc		C12	Max	10
Group 4	Alhydrogel + 13.6 µg F1-WV fusion protein days 0,30,sc		C12	Max	10
Group 5	Alhydrogel + 27.2 µg F1-WV fusion protein days 0, 30, sc		C12	Max	10
Group 6	Alhydrogel alone days 0, 30, sc		C12	Max	10 NC
Group 7	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc		CO92	100	10
Group 8	Alhydrogel alone, days 0, 30, sc		CO92	100	10 NC
Aerosol challenge					
Group 09	Alhydrogel alone, days 0, 30, sc		C12	50	10
Group 10	Alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc		C12	50	10
Group 11	Alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc		C12	Max	10
Group 12	Alhydrogel + 13.6 µg F1-WV fusion protein, days 0, 30, sc		C12	Max	10
Group 13	Alhydrogel + 27.2 µg F1-WV fusion protein days 0,30, sc		C12	Max	10
Group 14	Alhydrogel alone, days 0, 30, sc		C12	Max	10
Group 15	Alhydrogel + 13.6 µg F1-WV fusion protein days 0, 30, sc		CO92	100	10
Group 16	Alhydrogel alone days 0, 30, sc		CO92	100	10
Group 17	Alhydrogel + 10 µg F1 + 10 µg Mauro's V, days 0, 30, sc		C12	Max	10
Group 18	Greer plague vaccine, days 0, 30, sc		C12	Max	09 NC
Group 19	Alhydrogel alone, day 0, 30, sc <i>LIT 10/13 w2</i>		C12	Max	05
Group 20	ALH + 13.6 µg F1-WV fusion protein day 0, 30, sc --antibody response Measure titer at 1:14, 27,57, lung lavage day 57				10

Parrot/birds in AA-4 have 1.8 air exchanges/minute. This is the key when

	Measure titer at X , 14, 27, 57; lung lavage day 57	
Group 22	ALH + Mauro-V urea, 10 ug, day 0, 30, sc --antibody response	10
Group 23	ALH alone, day 0,30, sc	10
	Measure titer at X 14, 27,57; lung lavage day 57	
Group 24	Greer plague vaccine days 0, 30, sc	
	Measure titer at X , 14, 27, 57; lung lavage day 57 Chipped	05
Group 25	Alhydrogel + 10 μ g F1 + 10 μ g Mauro's V, days 0, 30, sc	07
	Measure titer at X , 14, 27, 57; lung lavage day 57, Mice rec'd 14Jun95	
		Total 232

Schedule

Groups 1-19

13Jun95 Arrival of Swiss Webster mice, female 7-8 wks, Harlan Sprague Dawley in AA-3 (Barrier)

24Jun95 Chipped with BioMedic Data Systems transponders, West

27Jun95 1st immunization, day 0, Zimmerman, West, Giunanzio, Archer, Anderson

25Jul95 2nd Immunization, day 28, ~~Yell, west, Zimmerman, Archer, Anderson~~

24Aug95 Bleed to determine prechallenge titers, day 58 5664 - 5847

08Sep95 Challenge by aerosol & sc routes, day 58, AA-3-~~5848-5899~~ serum # 6082 - 6157

28Sep95 Terminal bleed, day 28 pi, titrate spleens# 10020 - 10094 serum # 6082 - 6157
~~Zimmerman, Giunanzio, Archer, Anderson, Shambur~~

Group 20-23

13Jun95 Mice arrive

27Jun95 1st immunization, AA-3

11Jul95 Groups 20, 21, 22; Bleed, day 14, AA-3, SERUM# 4542-4593

26Jul95 Groups 20, 21, 22; Bleed, day 29, AA-3, SERUM# 4807-4888 GIA# 210, HALL

25Jul95 2nd immunization, day 28, new barrier, AR-5

01Sep95 Bleed, day 65, AA-3, Groups 24, 25 SERUM# 5848-5899 GIA# 210, HALL
 Groups 20, 21, 22, and 23, for serum titer & bronchial lavage #5900 - 5951

01Sep95 Group 23 for Spleen weights # 10010 - 10019
 ARCHER, DR PULLMAN

Chip numbers for all groups extra alhydrogel controls

No Chip GP1
 No Chip
 No Chip

200E2C4363/WV-001 GP2
 20103F1F72/WV-002
 1F73663C4C/WV-003
 1F29341D67/WV-004
 2001693A3C/WV-005
 2000183C0C/WV-006
 1F561F4725/WV-007
 1F5F027808/WV-008
 1F4B1D3346/WV-009
 20023A3371/WV-010

Plaque size in AA-4 for 1.8 g per ephelio/ female. This will be lot when

1F687D403F/WV-012
1F46376400/WV-013
2041670F29/WV-014
20024E4B45/WV-015
1F6136301B/WV-016
1F734C475B/WV-017
1F575E2C00/WV-018
200E317928/WV-019
1F5718640E/WV-020
1F22052119/WV-021 GP4
200B54374A/WV-022
1F76733B3D/WV-023
201D32642D/WV-024
1F757D303F/WV-025
200D150F2F/WV-026
203C120210/WV-027
200F585326/WV-028
1F667A2859/WV-029
1F56224D1C/WV-030
200B4A612A/WV-031 GP5
201D3B1B6D/WV-032
2041744C5F/WV-033
1F41111877/WV-034
20430C256C/WV-035
1F570E7408/WV-036
1F565A1120/WV-037 Missing chip *new 7F7A0E 7B4E*
1F735B6D26/WV-038
1F7F320828/WV-039
2003135E6C/WV-040
No Chip GP6
No Chip
1F73175007/WV-041 GP7
1F63711B72/WV-042
1F7C42653E/WV-043
200B622C47/WV-044
1F7F2E5D57/WV-045
1F66752C5A/WV-046
1F5E776329/WV-047
1F4E420B46/WV-048
200E4D364F/WV-049
203C1D2463/WV-050
No Chip GP8
No Chip
No Chip
No Chip

11/11/04 1.8 min exchange/insert. Then with 30% when

No Chip
No Chip
No Chip
No chip

2002537219/WV-051 GP9
201D31583A/WV-052
201D404B38/WV-053
200D22753C/WV-054
1F56211654/WV-055
1F5F154528/WV-056
200A195D60/WV-057
200C043917/WV-058
200142120B/WV-059
1F561F5814/WV-060
201D2D4650/WV-061 GP10
201D30276C/WV-062
1F73085B0B/WV-063
1F757A0A68/WV-064
200F353666/WV-065
203C6C2216/WV-066
200657087B/WV-067
20415E0E33/WV-068
1F501B492D/WV-069
200D7A4D0C/WV-070
1F58195B15/WV-071 GP11
1F7C414D57/WV-072
200A024E06/WV-073
200D34514E/WV-074
1F7F373873/WV-075
2018766969/WV-076
1F61110A65/WV-077
200B0C1732/WV-078
200D51136F/WV-079
1F657E7608/WV-080
20001F4D74/WV-081 GP12
20197C6467/WV-082
1F560B532D/WV-083
2018742331/WV-084
1F28782021/WV-085
203C0C5C3C/WV-086
1F402E787B/WV-087
200B386439/WV-088
200D7E7D58/WV-089
200666175D/WV-090
2019711D39/WV-091 GP12
20017C5211/WV-092
201B493C40/WV-093
1F4E1D383E/WV-094
200B691F4D/WV-095
1F63265C7C/WV-096
1F56147502/WV-097
2041632517/WV-098

weight & time of oscillating

34.2
29.0
34.4
41.9
29.8
37.5
28.5
28.3
34.9
37.4

$\frac{335.9}{10} = 33.6 \text{ gm avg. weight}$

Punkboy's pendulum on 911-4 has 1.8 sec. oscillation/minute. This is the best value.

1F3D4E5F60/WV-101 GP14

1F5A1M0031/WV-102

200F4D7200/WV-103

200F4D71D0/WV-104

200F4D5A84/WV-105

201D482F4F/WV-106

2008039A20/WV-107

1F7C7C4227/WV-108

2035422049/WV-109

201D2E2C89/WV-110

203A665100/WV-111 GP15

1F561E2E3F/WV-112

1F87015930/WV-113

1F5626184D/WV-114

200F31011F/WV-115

201E6C4313/WV-116

2018704C0C/WV-117

1F19690758/WV-118

200F136D51/WV-119

1F76132731/WV-120

201052314D/WV-131 GP16

1F5615492D/WV-132

1F73656128/WV-133

2001311618/WV-134

20104C6E16/WV-135

201C317E15/WV-136

200D48117A/WV-137

200046504A/WV-138

1F62746427/WV-139

201D483546/WV-140

201D2E2A6B/WV-141 GP17

200A05272A/WV-142

2000137756/WV-143

200C06410D/WV-144 - 1F6C7F3343

1F7D60453F/WV-145

2041580443/WV-146

1F75787C78/WV-147

1F20264457/WV-148

1F560D116D/WV-149

1F73671E69/WV-150

No chip GP18

No chip

2000037B62/WV-122 GP19

1F6679384A/WV-123

20093B4D4F/WV-125

0... last barcode in AA-4 has 1.8 mm spacing/minute. Take in the top when

1F1E01154D/WV-160

1F001E01154D/WV-161

1F001E01154D/WV-162

1F001E01154D/WV-163

1F001E01154D/WV-164

1F001E01154D/WV-165

1F001E01154D/WV-166 GP21

1F1E01154D/WV-167

200A112F16/WV-168

1F88244824/WV-169

200D2F170D/WV-170

20097A0857/WV-161

200C032829/WV-162

2010346339/WV-163

1F743C6948/WV-164

201F012818/WV-165

201D480C8F/WV-176 GP22

2035323B3E/WV-177

1F29476B06/WV-178

1F791B202D/WV-179

20415D784A/WV-180

203C137F12/WV-171

1F60390543/WV-172

1F76001259/WV-173

1F56035E2A/WV-174

20447C326E/WV-175

1F7C4C0217/WV-186 GP23

1F203F156D/WV-187

2001361415/WV-188

1F7C2B0733/WV-189

1F73571B7C/WV-190

1F6337794E/WV-181

200B5E4C2B/WV-182

1F60786524/WV-183

20024F226D/WV-184

1F56155B1B/WV-185

200F6B1551/WV-121 GP24

Print as 12/95, case unknown

1F2052056A/WV-124

2009221C19/WV-126

1F2A6A1E2F/WV-128

2008425640/WV-130

No Chip GP25

No Chip

REDACTED

Project: Recombinant F1-wholeV Fusion

Notebook #: 3739

Inoculum: *Yersinia pestis* strain, CO92

Route: sc

Dose-

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions.

Discard dead animals

Use scanner to check chip number of dead mice

Mark number of animals alive in each cage

re = mouse has been rechipped

See page 29

Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions
card dead animals

Card dead animals
→ seems to check

Scanner to check chip number of dead mice
etc number of animals alive in each cage

1) Number of animals alive in each cage
- mouse has been euthanized

= mouse has been rechipped

REDACTED	Project: Recombinant F1-wholeV Fusion					
Notebook #: 3739						
Inoculum: Yersinia pestis strain, CO92 OR C12						
Route: AEROSOL	Dose:	REDACTED	7-8wks	Vendor: Harlan Sprague Dawley	Sex: female	
REDACTED	Swiss Webster	A	REDACTED			
Day postinfection	8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30	1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29	1 2 3 4 5 6 7			
Group	LD50				Comments/Chip #	
15	CO92	BLACK	WHITE		203A555100/WV-111	
Alhydrogel 13.6ug F1-WV					1F561E2E3F/WV-112	
					1F57015930/WV-113	
					1F5626184D/WV-114	
					200F31011F/WV-115	
					201E6C4313/WV-116	
					2018704C0C/WV-117	
					1F19690758/WV-118	
					200F136D51/WV-119	
					1F76132731/WV-120	
16	CO92	BLACK	WHITE		201052014D/WV-181	
Alhydrogel alone					1F5613492D/WV-182	
					1F73656128/WV-183	
					2001311618/WV-184	
					20104C6E16/WV-185	
					201C317E15/WV-186	
					200D40117A/WV-187	
					200044594A/WV-188	
					1F02740427/WV-189	
					201D483546/WV-140	
17	C12	GREEN	WHITE		201D2E2A6B/WV-141	
Alhydrogel 10ug F1 10ug Mauro-V	Max	Black	Black		200A05272A/WV-142	
		Run 1	Run 2		2000137756/WV-143	
		Run 3	Run 4		1F6C7F3343/WV-144 re	
		Run 5	Run 6		1F7D60453F/WV-145	
		Run 7	Run 8		2041580443/WV-146	
		DIED	DURING		1F75787C78/WV-147 DIED	
		Run 9	Run 10		1F20264457/WV-148	
		Run 11	Run 12		1F560D116D/WV-149	
		Run 13	Run 14		1F73671E69/WV-150	
18	C12	Blue	White		No chip - DIED	
Greer Plague Vaccine	Max	Black	Black		No chip	
		Run 1	Run 2		No chip	
		Run 3	Run 4		No chip	
		Run 5	Run 6		No chip	
		Run 7	Run 8		No chip	
		Run 9	Run 10		No chip	
		Run 11	Run 12		No chip	
		Run 13	Run 14		No chip	
		Run 15	Run 16		No chip	
		Run 17	Run 18		No chip	
		Run 19	Run 20		No chip	
19	C12	Black	White		2000037802/WV-122	
Alhydrogel alone	Max	Run 1	Run 2		1F6679384A/WV-123	
		Run 3	Run 4		20099B4D4F/WV-125	
		Run 5	Run 6		201E073506/WV-127	
		Run 7	Run 8		WV-129 Missing chip	
		Run 9	Run 10		2D35406407	
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
20	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
21	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
22	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
23	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
24	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
25	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
26	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
27	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
28	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
29	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
30	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
31	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
32	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
33	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
34	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
35	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
36	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
37	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
38	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			
		Run 7	Run 8			
		Run 9	Run 10			
		Run 11	Run 12			
		Run 13	Run 14			
		Run 15	Run 16			
		Run 17	Run 18			
		Run 19	Run 20			
39	C12	Black	White			
Alhydrogel alone	Max	Run 1	Run 2			
		Run 3	Run 4			
		Run 5	Run 6			

File: F1-V, longterm Last update: REDACTED
 Protocol: B95-01
 Investigators: Anderson/Heath/Welkos/Friedlander

Background: F1-WV and F1 and V in combination have been shown to protect against challenges of CO92 and C12 with a two dose schedule (0 and 30). The long-term decay of the antibody response to the initial immunization and length of protection from a single immunization is currently not known.

Purpose: To examine the decay of the antibody response to an initial immunization, protection afford over time to an initial immunization to indicate the optimum time for the 2nd immunization for an aerosol challenge. Titers to F1 and V will be determined. Mice will be challenged with 50-100 LD₅₀ CO92, aerosol challenge.

In the below challenge groups, when protection falls to zero, the remaining groups will be booster and challenged 2 weeks post-boost.

Immunogens: EcF1s, Mauro's V, and F1-WV all essentially endotoxin free from Dr. Heath.

Mice: Swiss Webster (Hsd:ND4) female mice per group from Harlan Sprague Dawley, Inc. Indianapolis, Indiana (317) 894-7521.

Alhydrogel: 1.3%, from SuperFos. Batch # 2043, Expiration date None, 0.1857 mg of AL/dose

IMI-1000, Implantable micro identification transponders, BioMedic Data Systems, Inc., 255 W. Spring Valley Ave. Maywood, NJ 07607, Tel 201 587-8300.

Plague USP, Lot # 1128X1, Expiration Date REDACTED, Greer Laboratories, Inc. P.O Box 800 Lenoir, NC 28645-0800

Group	Treatment	Challenge Day	# of mice
1A	10μF1+20μgMauro-V	day0	14
1B			10
2A	30μgF1-WV	14	10
2B			10
3	Plague USP	14	10
4	Alhydrogel alone	14	10
5A	10μF1+20μgMauro-V	day0	42
5B			10
6A	30μgF1-WV	42	10
6B			10
7	Plague USP	42	10
8	Alhydrogel alone	42	10
9A	10μF1+20μgMauro-V	day0	98/119
9B			10
10A	30μgF1-WV	98/119	10
10B			10
11	Plague USP	98/119	10
12	Alhydrogel alone	98/119	10

serum # 6990-7039, pre challenge
 serum # 10146-10160, day 28 PI, pre challenge
 serum # 7820-7833, day 28 PI, pre challenge
 serum # 7676-7725, pre challenge
 serum # 10160-10191, day 30 PI
 serum # 8004-8035, day 30 PI, pre challenge
 serum # 8288-8337, pre challenge
 serum # 8591-8621, day 28 PI
 serum # 10259-10290, day 28 PI

13A	10 μ F1+20 μ gMauro-V	day0	10	10
13B			05	10
14A	30 μ gF1-WV		10	10
14B			05	10
15	Plague USP		10	10
16	Alhydrogel alone		10	10
17A	10 μ F1+20 μ gMauro-V	day0	10	10 serial bleeds on challenge days,
17B			05	
18A	30 μ gF1-WV		10	14, 42, 93, and _____
18B			05	day 14 serum # 7040 - 3027
19	Plague USP		10	# 7822 - 7833 7771 - 7819
20	Alhydrogel alone		10	42
			93	= 8400 - 8449
			Total 250	2000 = 8906 - 8955

Schedule

17Oct95 Mice arrive B412
 25Oct95 Mice chipped B412, Plumtree, Zimmerman
 31Oct95 Mice immunized day 0, Plumtree, Zimmerman
 07Nov95 Bleed 1st challenge group, day 7 ~~serum~~⁴
 14Nov95 1st challenge group, day 14, 100 LD₅₀ aerosol challenge, CO92
 14Nov95 Bleed serial bleed group, day 14
 05Dec95 Bleed 2nd challenge group, day 35
 12Dec95 2nd challenge group, day 42, 100 LD₅₀ aerosol challenge, CO92
 12Dec95 Bleed serial bleed group, day 42
 25Jan96 Bleed 3rd challenge group, day 96 *PLUMTREE, SAM BURN, 1/21/96*
 01Feb96 3rd challenge group, day 93, 100 LD₅₀ aerosol challenge, CO92
 01Feb96 Bleed serial bleed group, day 93
 ____95 Bleed 4th challenge group, day
 ____96 4th challenge group, day, 100 LD₅₀ aerosol challenge, CO92
 ____96 Bleed serial bleed group, day

Chips Numbers

22254D6722/LT-001 GP1A
22254B4164/LT-002
221D487E7E/LT-003
221D705617/LT-004
22213D0B42/LT-005
222122186F/LT-006
22223E1239/LT-007
2221493775/LT-008
221D670720/LT-009
22213D5220/LT-010
222233166C/LT-011 GP2A
2227765159/LT-012
221D6C477D/LT-013
2222485A77/LT-014
221A2B093D/LT-015
221D6B7917/LT-016
221D686E17/LT-017
2222411328/LT-018
22225C6018/LT-019
221D630D5E/LT-020
221D4E2460/LT-021 GP3

22280F622F/LT-022
221D570056/LT-023
221D552A40/LT-024
22223C1767/LT-025
2221534945/LT-026
22277D234D/LT-027
221D660734/LT-028
221B495727/LT-029
2225577D72/LT-030
221B45423B/LT-031 GP4
221B4C6576/LT-032
221D5B6077/LT-033
2222405171/LT-034
22277E167A/LT-035
221D682963/LT-036
222251294B/LT-037
221B464652/LT-038
221D605813/LT-039
221D762A24/LT-040
22224F0E15/LT-041 GP5A
22252A534C/LT-042
221D627877/LT-043
22214B1228/LT-044
2219346F3E/LT-045
221D4F3856/LT-046
2227545533/LT-047
22214E1004/LT-048
2222455747/LT-049
221B366372/LT-050
2227624147/LT-051 GP6A
22217E1A0D/LT-052
2228195A13/LT-053
2222585550/LT-054
2222390E64/LT-055
22214D1F3C/LT-056
2225405710/LT-057
221D713C0B/LT-058
22225B030A/LT-059
221D645754/LT-060
221B502C5D/LT-061 GP7
22276D2115/LT-062
222142305C/LT-063
22215D0B34/LT-064
2222375E5F/LT-065
2228166739/LT-066
221D6D0B4C/LT-067
2222576F77/LT-068
222155635F/LT-069
2222480418/LT-070
2221464260/LT-071 GP8
2222444E48/LT-072
221D72505E/LT-073
221D6D0524/LT-074
222243493A/LT-075

2221421752/LT-076
221D756C00/LT-077
2221586629/LT-078
22217A0417/LT-079
221D730574/LT-080
221D54560C/LT-081 GP9A
2221523113/LT-082
221D4E207A/LT-083
2225395749/LT-084
2221340347/LT-085
2221774E53/LT-086
2222576612/LT-087
22224E6F58/LT-088
221D563E57/LT-089
22212D0F34/LT-090
221D62392D/LT-091 GP10A
2222411F74/LT-092
221D4D6C2C/LT-093
221D554048/LT-094
2221473A36/LT-095
2221240C60/LT-096
22214F1224/LT-097
2221425212/LT-098
22223C390C/LT-099
2221414605/LT-100
221D6F1230/LT-101 GP11
2221305178/LT-102
2221770F09/LT-103
221D403C64/LT-104
2222355935/LT-105
221D480305/LT-106
2221633E37/LT-107
22254A6B01/LT-108
222147520A/LT-109
2221760056/LT-110
2222401108/LT-111 GP12
2222422877/LT-112
2227610A1C/LT-113
221D686D00/LT-114
221D703E2B/LT-115
221D671449/LT-116
22225C0824/LT-117
2221586D78/LT-118
22214C0710/LT-119
221D753C07/LT-120
2227712235/LT-121 GP13A
222241257B/LT-122
22224F1419/LT-123
221D744B1A/LT-124
22224E7477/LT-125
22224C4154/LT-126
221D66020D/LT-127
2221427332/LT-128
221D730F06/LT-129

221D524811/LT-131
221D524812/LT-132
221D524813/LT-133
221D524814/LT-134
221D524815/LT-135
221D524816/LT-136
222229562/B004/LT-137
2221754939/LT-138
221D480C51/LT-139
222153781B/LT-140
221D481476/LT-141 GP15
221D697718/LT-142
2228163110/LT-143
2222526107/LT-144
22217D4C57/LT-145
22222E5018/LT-146
221B2C4420/LT-147
222156603B/LT-148
2221522454/LT-149
22216B412F/LT-150
2221477A4F/LT-151 GP16
2222581133/LT-152
22223B4F59/LT-153
2221261D5A/LT-154
2221793120/LT-155
221D725567/LT-156
221D5E4642/LT-157
22212C640B/LT-158
22274C6575/LT-159
22252D2519/LT-160
221B310F2B/LT-161 GP17A
222172726A/LT-162
221D6E3F58/LT-163
2221321B14/LT-164
221B366F2E/LT-165
22225B0229/LT-166
221D6D4250/LT-167
22224E340E/LT-168
221D70795F/LT-169
22252F057D/LT-170
221D747D49/LT-171 GP18A
2221440747/LT-172
22225A5D71/LT-173
221D6D464A/LT-174
2221421E37/LT-175
22275A7201/LT-176
221D750C7A/LT-177
22225A7B5C/LT-178
2227600C26/LT-179
222244472D/LT-180
22224B496D/LT-181 GP19
2225385C0C/LT-182
221D524871/LT-183

221D742C6D/LT-184
221B471829/LT-185
221D59352E/LT-186
2222476C0C/LT-187
2222511C0F/LT-188
22277F7C66/LT-189
221D4B422C/LT-190
221D6B4536/LT-191 GP20
2221710D56/LT-192
22281C2918/LT-193
221D5C6E60/LT-194
2221662F72/LT-195
221D5E3068/LT-196
22223A6E6D/LT-197
22277C0B1C/LT-198
221D6C3C28/LT-199
2228045804/LT-200
2221463F1B/LT-201 GP1B
22213F0074/LT-202
22215A2E71/LT-203
22223B1136/LT-204
22215A7309/LT-205
221D6B4C53/LT-206 GP2B
221D49253C/LT-207
221D710C76/LT-208
22273F017C/LT-209
22217E5E6E/LT-210
221D494F34/LT-211 GP5B
22217B022D/LT-212
2221386837/LT-213
2228057813/LT-214
222142434F/LT-215
2222323813/LT-216 GP6B
222121405D/LT-217
2221755864/LT-218
2225527970/LT-219
221D56403B/LT-220
221D681542/LT-221 GP9B
22280E312B/LT-222
221D584D78/LT-223
221D4C4067/LT-224
22212A2966/LT-225
222124374C/LT-226 GP10B
2222586303/LT-227
2222371808/LT-228
2222443D5B/LT-229
22213B707C/LT-230
221D737744/LT-231 GP13B
2221313035/LT-232
2222392702/LT-233
221D73716A/LT-234
221D527D35/LT-235
221B2E7B71/LT-236 GP14B
221D484645/LT-237

221A422E07/LT-230
221A422E08/LT-230
221A422E09/LT-230
221A422E09/LT-241 GP17B
221A422E09/LT-242
221A422E09/LT-243
221A422E09/LT-244
221A422E09/LT-245
1F7E000477/LT-246 GP18B
2222408E04/LT-247 - 695 CHIP, RESPONSED 1F7E1B695F
2221470734/LT-248
22254A3C0B/LT-249
221A422E06/LT-250
1F72777B7D/TESTCHIP

REDACTED		Project: Active F1-WV, F1+V Longterm study		DAY 14 POST IMMUNIZATION			
Notebook #: 3739							
neculum: Yersinia pestis strain, CO92		6030 82-108					
Route: AEROSOL Dose: as shown below							
REDACTED	Wiss Webster	Arr	REDACTED	7-8wks	Vendor: Harlan Sprague Dawley	Sex: female	
Day postinfection	0	1	2	3	4	5	6
Group	Run	7	8	9	10	11	12
1A	CO92	13	14	15	16	17	18
10ug F1+ 20ug nauro-v	19	20	21	22	23	24	25
1.6	26	27	28	29	30	1	2
10ug F1+ 20ug nauro-v	3	4	5	6	7	8	9
1.6	10	11	12	13	14	15	16
10ug F1+ 20ug nauro-v	17	18	19	20	21	22	23
1.6	24	25	26	27	28	29	1
1B	CO92	2	3	4	5	6	7
10ug F1+ 20ug nauro-v	8	9	10	11	12	13	14
1.6	15	16	17	18	19	20	21
10ug F1+ 20ug nauro-v	22	23	24	25	26	27	28
1.6	29	30	1	2	3	4	5
2A	CO92	1	2	3	4	5	6
10ug F1- WV	7	8	9	10	11	12	13
2.6	14	15	16	17	18	19	20
10ug F1- WV	21	22	23	24	25	26	27
2.6	28	29	30	1	2	3	4
2B	CO92	1	2	3	4	5	6
0ug F1- WV	7	8	9	10	11	12	13
2.6	14	15	16	17	18	19	20
0ug F1- WV	21	22	23	24	25	26	27
2.6	28	29	30	1	2	3	4
3	CO92	1	2	3	4	5	6
lague USP inter	7	8	9	10	11	12	13
3.6	14	15	16	17	18	19	20
lague USP inter	21	22	23	24	25	26	27
3.6	28	29	30	1	2	3	4
4	CO92	1	2	3	4	5	6
hydrogel cone	7	8	9	10	11	12	13
3.6	14	15	16	17	18	19	20
hydrogel cone	21	22	23	24	25	26	27
3.6	28	29	30	1	2	3	4

or Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

iscard dead animals

heck chip number of dead mice with scanner

lark number of animals alive in each cage

OST after Chip # means the chip has fallen out

HEAD COLOR
BLACK RUN 6
RED RUN 7

C HIR HANGING OUT

88
Exhibit GA11

File: Alhydrogel concentration

Last updated: **REDACTED**

Anderson/Heath/Welkos/Friedlander

Background. The allowable Al content in a human vaccine is 0.85 mg/dose as determined by assay (21 CFR 610.15(1)). However, the lowest possible dose of Al should be used which maintains an adequate adjuvant response with the EcF1c and V immunogens. A dose response for Alhydrogel has not been done with a combination of F1 and V. Therefore this experiment will examine a range of concentrations of AL which will be used with a constant amount of EcF1c and V. EcF1c (60 EU/ml) and V-His tag (preparation are essentially endotoxin free. The level of endotoxin in the thrombin treated V preparation without the His tag is 49 EU/ml.

Compare the antibody response to V with F1-WV protein with and without alhydrogel. When F1 + V is used to immunize without alhydrogel, there was not antibody response to V. This will be done with F1-WV in order to determine whether F1 is contributing anything to protection with the F1-WV protein. F1-WV purified by Ni++ and ran through a Sartorius Q15 filter using 10mMtris, pH 7.6, 0.5mM EDTA + 0.5 MNaCl for elution. See Heath's note of 5-6 Dec. Endotoxin _____ EU/ml.

Mice: Swiss Webster (Hsd:ND4) female mice from Harlan Sprague Dawley, Inc. Indianapolis, Indiana (317) 894-7521.

1.3% Alhydrogel (Aluminium Hydroxide Gel Adjuvant): Al_2O_3 (1.3%) equivalent to $Al(OH)_3$ (2.0%), from SuperFos Biosector a/s, Frydenundsvej 30, DK-2950 Vedbaek, Denmark. Batch # 2043, Expiration date None: U.S. supplier - Accurate Chemical & Scientific Corp, 300 Shames Drive, Westbury, NY 11590, Tel (516) 333-2221, Fax (516) 997-4948.

Al = 13 O = 8 H = 1, $Al(OH)_3$ = 40 molecular weight

Current procedure for adsorption of F1 and V to Alhydrogel: 1.0ml Alhydrogel brought to 7.0 ml
2% $Al(OH)_3$ = 20 mg/ml

$20mg/ml)(1.0ml) = (x)(7.0ml$ final volume) $x = 2.857$ mg $Al(OH)_3/ml$

$(2.857mg/ml)(0.2ml$ dose) = 0.57142 mg $Al(OH)_3$

AL is 0.325% of $Al(OH)_3$

0.235% of 2.857mg = 0.1857 mg of AL/0.2ml dose which the mouse receives

Current dose of Al which has been used through out the mouse experiments is 0.1857mg. Try two other doses each 75% of the former GP1 = 0.1857mg (standard amount of AL), GP2 = 0.1393mg, GP3 = 0.1045mg

IMI-1000, Implantable micro identification transponders, BioMedic Data Systems, Inc., 255 W. Spring Valley Ave. Maywood, NJ 07607, Tel 201 587-8300.

Plague USP, Lot # 1128X1, Expiration Date **REDACTED**, Greer Laboratories, Inc. P.O Box 800 Lenoir, NC 28645-0800,

Groups	Treatment	V-HIS Tag	Strain	#Mice
000A	30 μ gF1-WV, μ g AL	Yes	CO92	10
000B	30 μ gF1-WV, μ g AL	Yes	CO92	05
00A	30 μ gF1-WV+0.19AI	Yes	CO92	10
00B	30 μ gF1-WV+0.19AI	Yes	CO92	05
0A	10 μ gF1+20 μ gV+0.19AI	No	CO92	10
0B	10 μ gF1+20 μ gV+0.19AI	No	CO92	10

1A	10µgF1+20µgV+0.19Al	Yes	CO92	10
1B	10µgF1+20µgV+0.19Al	Yes	CO92	10
2A	10µgF1+20µgV+0.14Al	Yes	CO92	10
2B	10µgF1+20µgV+0.14Al	Yes	CO92	10
3A	10µgF1+20µgV+0.10Al	Yes	CO92	10
3B	10µgF1+20µgV+0.10Al	Yes	CO92	10
4	0.19Alhydrogel		CO92	10
5	0.19Alhydrogel		CO92	10
6	0.14Alhydrogel		CO92	10
7	0.10Alhydrogel		CO92	10
8	10µgF1+20µgV, No Alh	Yes	CO92	10
9	10µgF1+20µgV, No Alh	Yes	CO92	10
10	Plague USP (Greer)sc		CO92	10
11	Plague USP (Greer)sc		CO92	10
12	No treatment		CO92	10
13	Plague USP (Greer) im		CO92	10
			Total	210

Total 210

Date	Procedure
09Nov95	Arrival of mice in B412 at 7-8 weeks of age
16Nov95	Chipped mice in B412, SGT Zimmerman/Plumtree
06Dec95	Day 0, immunization Anderson/Shamblin
12Jan96	Day37, bleed prior to challenge, serum # 8045-8254 PLUMTREE, ANDERSON, SHAMBLIN
19Jan96	Day44, challenge ANDERSON, WILKINSON, + B10T, 1 st 3 RUNS
15Feb96	Day 28 pi, terminal bleed, spleen removal serum # spleen #

Chip numbers

221D682232/ALH-001	GP000A
2225353065/ALH-002	
222543363A/ALH-003	
2227471E04/ALH-004	
2221462517/ALH-005	
221D63634C/ALH-006	
221D645F12/ALH-007	
221B2F0113/ALH-008	
222537362C/ALH-009	
221B4D572B/ALH-010	
221D3A1A63/ALH-011	GP000B
2227593C63/ALH-012	
22213E5370/ALH-013	
2228111E27/ALH-014	
221B4D060F/ALH-015	
2221291C51/ALH-021	GP00A
22217A1916/ALH-022	
22215A4825/ALH-023	
22213D0C4F/ALH-024	
2221661921/ALH-025	
2221501E3C/ALH-026	
221D62561C/ALH-027	
2221796313/ALH-028	
22216A4521/ALH-029	
221D5B654E/ALH-030	
221D5B165D/ALH-031	GP00B
2225535315/ALH-032	

See pages 104-107

From USMEL 10/10/2010

2222382716/ALH-033
221D64205D/ALH-034
2221361744/ALH-035
221B3B756D/ALH-041 GP0A
2222505C62/ALH-042
221D355F0B/ALH-043
2221393824/ALH-044
2221260562/ALH-045
2221750728/ALH-046
2221710C75/ALH-047
22254A533A/ALH-048
221D48425F/ALH-049
222547047F/ALH-050
221B431E0B/ALH-051 GP0B
222141267F/ALH-052
2225447977/ALH-053
22217C5904/ALH-054
22276E4F74/ALH-055
2222513264/ALH-056
221B331A0B/ALH-057
22223A2C20/ALH-058
2225301E16/ALH-059
2222464F0C/ALH-060
2225525E7E/ALH-061 GP1A
22214B6521/ALH-062
2225427F32/ALH-063
221B2E1A28/ALH-064
22277F2A4F/ALH-065
221B381564/ALH-066
22276C4058/ALH-067
221B365521/ALH-068
2221483A1E/ALH-069
221B2F4049/ALH-070
2222490E7E/ALH-071 GP1B
22216B6702/ALH-072
22281D7640/ALH-073
221D74327B/ALH-074
221B36013C/ALH-075
2227177522/ALH-076
221B560D16/ALH-077
222734024F/ALH-078
221B463462/ALH-079
221B463656/ALH-080
22252D6D26/ALH-081 GP2A
2225416D0B/ALH-082
221B360D60/ALH-083
22215C0152/ALH-084
222550400F/ALH-085
221B496E3F/ALH-086
2221353A4B/ALH-087
221D542860/ALH-088
2225376005/ALH-089
221B3A5805/ALH-090
221D74480D/ALH-091 GP2B

See page 104-107

2225315130/ALH-093
2221630F69/ALH-094
221D615D3E/ALH-095
2225574367/ALH-096
221B440F29/ALH-097
222176302B/ALH-098
22212E212C/ALH-099
22273B1603/ALH-100
22274C0115/ALH-101 GP3A
222178537A/ALH-102
2225447B43/ALH-103
2221775C19/ALH-104
2227351528/ALH-105
22271B745A/ALH-106
2221525A38/ALH-107
2225546B51/ALH-108
2225316828/ALH-109
222243260B/ALH-110
2221694D14/ALH-111 GP3B
22277D2440/ALH-112
2225407A6C/ALH-113
2221445B1C/ALH-114
221D6E5550/ALH-115
2227514F27/ALH-116
22214B6D67/ALH-117
22254F0206/ALH-118
221B3B1F65/ALH-119
222543090C/ALH-120
2225402D66/ALH-121 GP4
2228015179/ALH-122
2225343914/ALH-123
222529444C/ALH-124
22271B7357/ALH-125
222732682C/ALH-126
2221501931/ALH-127
22196E7321/ALH-128
22280F5F2D/ALH-129
222551747C/ALH-130
2222464E2F/ALH-131 GP5
22272E117A/ALH-132
22225C6636/ALH-133
22254E3756/ALH-134
22253D1205/ALH-135
22216A047B/ALH-136
22216C346D/ALH-137
2225572C56/ALH-138
221B597D3A/ALH-139
221B3D2F73/ALH-140
221D6A064C/ALH-141 GP6
22254C574B/ALH-142
22224D0523/ALH-143
2225561B32/ALH-144
2227475E7D/ALH-145

*From VSMW 100 milton
of Dan Heath*

104-107

*from system monitor
of Dan D. and*

2221702/33/ALH-146
221D504A22/ALH-147
221B330124/ALH-148
22215C754C/ALH-149
22253B326D/ALH-150
2222586C48/ALH-151 GP7
222232667C/ALH-152
2228452429/ALH-153
2222306C69/ALH-154
222836351D/ALH-155
222779775C/ALH-156
221B377621/ALH-157
2222544F07/ALH-158
2221734C6B/ALH-159
2222442E32/ALH-160
2222575041/ALH-161 GP8
2222511267/ALH-162
2221516F0F/ALH-163
221B2F761A/ALH-164
22276B7F11/ALH-165
2222370F7B/ALH-166
222164005D/ALH-167
221D705734/ALH-168
221B465E6A/ALH-169
22224C3E1B/ALH-170
221D592473/ALH-171 GP9
22280E6F44/ALH-172
22223D0B51/ALH-173
2225376F4E/ALH-174
2221704167/ALH-175
22224D1769/ALH-176
2225312134/ALH-177
221D611735/ALH-178
222245080B/ALH-179
221B39427A/ALH-180
2222490F5D/ALH-181 GP10
22271D2B7D/ALH-182
2227337D7F/ALH-183
22225C0D1D/ALH-184
221D4C6901/ALH-185
22277B6E20/ALH-186
2221690A60/ALH-187
222846320A/ALH-188
222161387E/ALH-189
221B30696B/ALH-190
22282E3D60/ALH-191 GP11
2221490E6D/ALH-192
2221627F79/ALH-193
2222321175/ALH-194
2221641439/ALH-195
2225443B3A/ALH-196
2222585764/ALH-197
22213B7E14/ALH-198
222160194A/ALH-199

REDACTED

2221490005/ALH-200
22213D1B3C/ALH-201 GP12
2222523865/ALH-202
2221707A4B/ALH-203
221B41083C/ALH-204
222837504A/ALH-205
22273A4C62/ALH-206
22216B1128/ALH-207
221B511B39/ALH-208
2222557D5A/ALH-209
221B432713/ALH-210 GP13
221D73342A/ALH-016
221D492B54/ALH-017
2228145E46/ALH-018
221D763028/ALH-019
2221543C07/ALH-020
22225A6150/ALH-036
2221534A52/ALH-037
221D73450D/ALH-038
22276D5C6E/ALH-039
2227557710/ALH-040

See page 104-107

REDACTED

From USAMRIID notebook
of Dave Heath

22211/11/2004/ALH-203
221B41083C/ALH-204
222837504A/ALH-205
22273A4C62/ALH-206
20016R110R/ALH-207

REDACTED

SET UP ABSORPTIONS FOR LTC ANDERSON:

GAPS 000A + 000B used PI-V that had been purified by Ni^{2+} & run through a Sartorius Q15 filter using 10 mM Tris, pH 7.6 0.5 mM EDTA & 0.5 M NaCl for elution. Buffer exchanged this using 10 mM Tris, pH 7.6 & 0.5 mM EDTA giving a conclusion. Did BCA assay for protein conc = 690 $\mu\text{g}/\text{mL}$ and 1.5 μL total volume

so used 450 μg in 3 mL PBS = 652 μL of PI-V ($\approx 690 \mu\text{g}/\text{mL}$)

652 μL PI-V + 2.348 mL PBS

GAPS 00A same PI-V as above @ 690 $\mu\text{g}/\text{mL}$
00B
- add 652 μL of PI-V to 428 μL ALYDROGEL + ABSORBED
PON 2 hr @ 4°C
- spin tube @ 2000 rpm for 5 min. & removed
2 100 μL aliquots to check for protein conc on BCA
assay (see BCA assay results)
- added PBS to absorbed PI-V to 3 mL

PS 0A + 0B Took PI capsule extract @ 60 $\mu\text{L}/\text{mL}$ = conc of 705 $\mu\text{g}/\text{mL}$
+ added 496 μL of this to 1 mL ALYDROGEL + 137 μL of
thrombin treated Vaseline (5.1 $\mu\text{g}/\text{mL}$) = 496 $\mu\text{g}/\text{mL}$
this is for 10 $\mu\text{g}/200 \mu\text{L}$ dose of PI + 20 $\mu\text{g}/200 \mu\text{L}$ dose V
in a final volume of 7 mL
496 μL of PI = 350 μg 137 μL V = 700 μg V
496 μL PI + 137 μL thrombin V + 367 μL PBS + 1 mL ALYDROGEL

REDACTED	(cont)		
GRPS 1A + 1B	10 μg F1 + 20 μg hV in 7 ml final volume $hV = 1.3 \text{ EU/ml}$ 496 μl F1 (705 μg/ml) + 129 μl hV (5.5 μg/ml) + 375 μl PBS + 1 ml ACHYDROCOR, Rock OV @ 40°C - Spin 2000 rpm, 5 min. remove 2 μl aliquots for BCA assay of absorption - Then added PBS to 7 ml final volume		
GRPS 2A + 2B	Same as 1A + 1B above but used 750 μl ACHYDROCOR + additional 250 μl PBS		
GRPS 3A + 3B	Same as 1A + 1B above but used 562.5 μl ACHYDROCOR + 437.5 μl PBS to equal 1 ml then same as 1A + 1B		
GRP 4 + 5	Added 1 ml ACHYDROCOR to 1 ml PBS + Rocked OV. Then added PBS to 7 ml final volume		
GRPS 6	Added 750 μl ACHYDROCOR + 250 μl PBS + 1 ml PBS + rocked OV @ 40°C. Then added PBS to 7 ml final volume.		
GRP 7	Added 562.5 μl ACHYDROCOR + 437.5 μl PBS then 1 ml PBS + Rocked OV @ 40°C - added PBS to 7 ml final volume		
GRP 8 + 9	Added 496 μl F1 + 129 μl hV + 1.375 ml PBS + Rocked OV. Then added PBS to 7 ml final volume		

REDACTED

BSA assay results:

BSA 1:10	.181	.157	PI + hV, 14 (2A+2B)	PI + hV, 19 undil (1A+1B)
1:20	.079	.086	#1 #2	#1 #2
1:40	.035	.041	.001	-.007 -.001
1:80	.012	.015		
			PI + hV (3A+3B), 10	PI + TV, 19 undil (0A+0B)
			#1 #2	#1 #2
			-.003 -.005	-.003 -.002
			PI-V purified, 19 (004+008)	
			.004	
			.003	

REDACTED

Sartorius Q15 experiment

- had 5ml of FIV C ~ 2mg/ml & did Q15 exp. as follows:

- 1) equilibrated Q15 in 15ml 10mM Tris (pH 7.6), 0.5mM EDTA,
- 2) applied 4.5ml of FIV in 10mM Tris, 0.5mM EDTA pH 7.6 to filter (collect "FLOW THRU")
- 3) washed Q15 in 10ml Tris-EDTA equilibration buffer & collected as "wash"
- 4) eluted in 12ml Tris, EDTA + 100mM NaCl & collected
- 5) " " " " " + 200mM NaCl " "
- 6) " " " " " + 300mM NaCl " "
- 7) " " " " " + 400mM NaCl " "
- 8) " " " " " + 500mM NaCl " "

1.00 ml... 0.100 collected each (test 1a)

Exhibit GA13

REDACTED	Project: F1+V alhydrogel concentrations, V w&wo His tag, F1-WV w&wo alhydrogel			
Notebook #: 3739				
Inoculum: <i>Yersinia pestis</i> strain, CO92				
Route: aerosol	Dose: as shown below		LD ₅₀ 190-304	
REDACTED	Swiss Webster	Arr.	REDACTED	7-8wks
Day	19 20 21 22 23 24	REDACTED	29 30 31	Vendor: Harlan Sprague Dawley
Day postinfection	C 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17	1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17	Sex: female	
Group	LD ₅₀			Comments/Chip #
000A F1-WV No alhydr 30ug	CO92 F7P			221D682232/ALH-001 222535065/ALH-002 222649963/ALH-003 2227471E04/ALH-004 2221462517/ALH-005 221D63634C/ALH-006 221D645F12/ALH-007 221B2F0113/ALH-008 222537362C/ALH-009 221B4D572B/ALH-010
000B F1-WV No alhydr 30ug	CO92			221D3A1A63/ALH-011 2227593C63/ALH-012 22213E5370/ALH-013 2228111E27/ALH-014 221B4D060F/ALH-015
00A F1-WV Alhydrogel 30ug	CO92			xxxxxx xxxxxx xxxxxx xxxxxx xxxxxx 2221291C51/ALH-021 22217A1916/ALH-022 22215A4825/ALH-023 22213D0C4F/ALH-024 2221661921/ALH-025 2221501E3C/ALH-026 221D62561C/ALH-027 2221796313/ALH-028 22216A4521/ALH-029 221D5B654E/ALH-030
00B F1-WV Alhydrogel 30ug	CO92			221D5B165D/ALH-031 2225535315/ALH-032 22223B2716/ALH-033 221D64205D/ALH-034 2221361744/ALH-035
0A 10F1+20V 0.19 Alh No His tag	CO92			221B3B756D/ALH-041 2222505C62/ALH-042 221D355F0B/ALH-043 2221393824/ALH-044 2221260562/ALH-045 2221750728/ALH-046 2221710C75/ALH-047 22254A533A/ALH-048 221D48425F/ALH-049 222547047F/ALH-050
0B 10F1+20V 0.19 Alh No His tag	CO92			221B431E0B/ALH-051 222141267F/ALH-052 2225447977/ALH-053 22217C5904/ALH-054 22276E4F74/ALH-055 2222513264/ALH-056 221B331A0B/ALH-057 22223A2C20/ALH-058 2225301E16/ALH-059 2222464F0C/ALH-060

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Check chip number of dead mice with scanner

F7P - MOUSE CAME OUT OF IT'S MICE PURPLE CHALLNGE

Mark number of animals alive in each cage

LOST after Chip # means the chip has fallen out

REDACTED

Project: F1+V alhydrogel concentrations, V w&wo His tag, F1-WV w&wo alhydrogel

Notebook #: 3739

Inoculum: Yersinia pestis strain, CO92

Route: aerosol Dose: as shown below

600 190-304

REDACTED		Swiss Webster		Arr	REDACTED		7-8wks		Vendor: Harlan Sprague Dawley												Sex: female											
REDACTED	Day	19	20	21	22	23	24	29	30	31	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	Comments/Chip #				
Day postinfection		0	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	17	18	19	20	21	22	23	24	25	26	27	28	29	
Group	LD50																															
4 0.19 Alh control	CO92	1	1	1	D																											2225402D66/ALH-121
		1	1	1	D																										2228015179/ALH-122	
		1	1	1	D																									2225343914/ALH-123		
		1	1	1	D																									222529444C/ALH-124		
		1	1	1	D																									22271B7357/ALH-125		
		1	1	1	D																									222732682C/ALH-126		
		1	1	1	D																									2221501931/ALH-127		
		1	1	1	D																									22196E7321/ALH-128		
		1	1	1	D																									22280E5E2D/ALH-129		
		1	1	1	D																									2225517470/ALH-130		
5 0.19 Alh control	CO92	1	1	1	D																									2222464E2F/ALH-131		
		1	1	1	D																									22272E117A/ALH-132		
		1	1	1	D																									22225C66367A/ALH-133		
		1	1	1	D																									22264E3756/ALH-134		
		1	1	1	D																									22253D1205/ALH-135		
		1	1	1	D																									22216A047B/ALH-136		
		1	1	1	D																									22216C346D/ALH-137		
		1	1	1	D																									2225572C56/ALH-138		
		1	1	1	D																									221959798A/ALH-139		
		1	1	1	D																									221B3D2F73/ALH-140		
6 0.14 Alh control	CO92	1	1	1	D																									221B6A064C/ALH-141		
		1	1	1	D																									22254C574B/ALH-142		
		1	1	1	D																									22224D0523/ALH-143		
		1	1	1	D																									2225561B32/ALH-144		
		1	1	1	D																									2227475E7D/ALH-145		
		1	1	1	D																									2221702733/ALH-146		
		1	1	1	D																									221D504A22/ALH-147		
		1	1	1	D																									221B330124/ALH-148		
		1	1	1	D																									22215C754G/ALH-149		
		1	1	1	D																									22253B326D/ALH-150		
7 0.10 Alh control	CO92	1	1	1	D																									2222586C48/ALH-151		
		1	1	1	D																									222232667C/ALH-152		
		1	1	1	D																									2228452429/ALH-153		
		1	1	1	D																									2222306C69/ALH-154		
		1	1	1	D																									222836351D/ALH-155		
		1	1	1	D																									222779775C/ALH-156		
		1	1	1	D																									2219977621/ALH-157		
		1	1	1	D																									2222544F07/ALH-158		
		1	1	1	D																									2221734C6B/ALH-159		
		1	1	1	D																									2222442E32/ALH-160		
8 10F1+20V No Alh His tag	CO92	1	1	1	D																									2222575041/ALH-161		
		1	1	1	D																									2222511267/ALH-162		
		1	1	1	D																									2221516F0F/ALH-163		
		1	1	1	D																									221B2F761A/ALH-164		
		1	1	1	D																									22276B7F11/ALH-165		
		1	1	1	D																									2222370P7B/ALH-166		
		1	1	1	D																									222164005D/ALH-167		
		1	1	1	D																									221D705734/ALH-168		
		1	1	1	D																									221B465E6A/ALH-169		
9 10F1+20V No Alh His tag	CO92	1	1	1	D																									22224C3E1B/ALH-170		
		1	1	1	D																									221D592473/ALH-171		
		1	1	1	D																									22280E6F44/ALH-172		
		1	1	1	D																									22223D0B51/ALH-173		
		1	1	1	D																									2222576F4E/ALH-174		
		1	1	1	D																									2221704167/ALH-175		
		1	1	1	D																									22224D1760/ALH-176		
		1	1	1	D																									2223312134/ALH-177		
		1	1	1	D																									221D611735/ALH-178		
		1	1	1	D																									222245080B/ALH-179		
		1	1	1	D																									221B39427A/ALH-180		

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Check chip number of dead mice with scanner

Mark number of animals alive in each cage

LOST after Chip # means the chip has fallen out

REDACTED

Project: F1+V alhydrogel concentrations, V w&wo His tag, F1-WV w&wo alhydrogel

Notre Dame 3738

Yersinia pestis strain, CO92

FluMist: aerosol Dose: as shown below

~~2050 190-304~~

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions.

Discard dead animals

Check chip number of dead mice with scanner

Mark number of animals alive in each cage

Mark number of animals alive in each cage
LOST after Chip # means the chip has fallen out

July 1988

Exhibit GA14

ELISA Summary: Protocol: F1-V Long Term (Day 7)

Plate	Serum	Group	Treatment	V ELISA DATE: 30 JAN 96	F1 ELISA DATE: 25 JAN 96
1A	6990	GP1A10ugF1	+200ugMaurV	320	485
1B	6991	GP1A10ugF1	+200ugMaurV	640	1,280
2A	6992	GP1A10ugF1	+200ugMaurV	2,560	10,240
2B	6993	GP1A10ugF1	+200ugMaurV	640	2,560
3A	6994	GP1A10ugF1	+200ugMaurV	640	2,560
3B	6995	GP1A10ugF1	+200ugMaurV	320	2,560
4A	6996	GP1A10ugF1	+200ugMaurV	320	2,560
4B	6997	GP1A10ugF1	+200ugMaurV	320	2,560
5A	6998	GP1A10ugF1	+200ugMaurV	640	5,120
5B	6999	GP1A10ugF1	+200ugMaurV	1,280	5,120
21A	7030	GP1B10ugF1	+200ugMaurV	320	1,280
21B	7031	GP1B10ugF1	+200ugMaurV	320	320
22A	7032	GP1B10ugF1	+200ugMaurV	320	5,120
22B	7033	GP1B10ugF1	+200ugMaurV	320	2,560
23A	7034	GP1B10ugF1	+200ugMaurV	320	5,120
6A	7000	GP2A 30ugF1-WV		320	557
6B	7001	GP2A 30ugF1-WV		640	1,280
7A	7002	GP2A 30ugF1-WV		640	5,120
7B	7003	GP2A 30ugF1-WV		2,560	
8A	7004	GP2A 30ugF1-WV		640	5,120
8B	7005	GP2A 30ugF1-WV		640	1,280
9A	7006	GP2A 30ugF1-WV		320	2,560
9B	7007	GP2A 30ugF1-WV		320	5,120
10A	7008	GP2A 30ugF1-WV		1,280	5,120
10B	7009	GP2A 30ugF1-WV		640	5,120
23B	7035	GP2B 30ugF1-WV		640	10,240
24A	7036	GP2B 30ugF1-WV		320	5,120
24B	7037	GP2B 30ugF1-WV		320	10,240
25A	7038	GP2B 30ugF1-WV		320	2,560
25B	7039	GP2B 30ugF1-WV		640	320
11A	7010	GP3 PlaqueUSP		320	343
11B	7011	GP3 PlaqueUSP		320	5,120
12A	7012	GP3 PlaqueUSP		320	2,560
12B	7013	GP3 PlaqueUSP		640	
13A	7014	GP3 PlaqueUSP		320	5,120
13B	7015	GP3 PlaqueUSP		640	640
14A	7016	GP3 PlaqueUSP		320	2,560
14B	7017	GP3 PlaqueUSP		320	2,560
15A	7018	GP3 PlaqueUSP		320	2,560
15B	7019	GP3 PlaqueUSP		320	5,120
16A	7020	GP4 alhydro alone		320	343
16B	7021	GP4 alhydro alone		320	320
17A	7022	GP4 alhydro alone		320	320
17B	7023	GP4 alhydro alone		320	320
18A	7024	GP4 alhydro alone		320	320
18B	7025	GP4 alhydro alone		320	320
19A	7026	GP4 alhydro alone		320	320
19B	7027	GP4 alhydro alone		640	320
20A	7028	GP4 alhydro alone		320	320
20B	7029	GP4 alhydro alone		320	320
33B	F/V POOL (+ CONTROL)			327,680	81,920

Protocol: F1-V Long Term (Day 35)

Plate	Serum	Group	Treatment	V ELISA DATE: 7 FEB 96	F1 ELISA DATE: 6 FEB 96
1A	7676	GP5A10ugF1	20ugVALH	1,310,720	1,040,319
1B	7677	GP5A10ugF1	20ugVALH	1,310,720	40,960
2A	7678	GP5A10ugF1	20ugVALH	1,310,720	40,960
2B	7679	GP5A10ugF1	20ugVALH	1,310,720	40,960
3A	7680	GP5A10ugF1	20ugVALH	1,310,720	20,480
3B	7681	GP5A10ugF1	20ugVALH	327,680	40,960
4A	7682	GP5A10ugF1	20ugVALH	1,310,720	20,480
4B	7683	GP5A10ugF1	20ugVALH	1,310,720	20,480
5A	7684	GP5A10ugF1	20ugVALH	1,310,720	40,960
5B	7685	GP5A10ugF1	20ugVALH	1,310,720	81,920
6A	7686	GP5B10ugF1	20ugVALH	1,310,720	10,240
6B	7687	GP5B10ugF1	20ugVALH	1,310,720	10,240
7A	7688	GP5B10ugF1	20ugVALH	1,310,720	20,480
7B	7689	GP5B10ugF1	20ugVALH	1,310,720	10,240
8A	7690	GP5B10ugF1	20ugVALH	163,840	20,480
8B	7691	GP6A ALH+	30ugF1-WV	655,380	948,482
9A	7692	GP6A ALH+	30ugF1-WV	1,310,720	5,120
9B	7693	GP6A ALH+	30ugF1-WV	655,380	20,480
10A	7694	GP6A ALH+	30ugF1-WV	1,310,720	10,240
10B	7695	GP6A ALH+	30ugF1-WV	1,310,720	10,240
11A	7696	GP6A ALH+	30ugF1-WV	163,840	2,560
11B	7697	GP6A ALH+	30ugF1-WV	655,380	2,560
12A	7698	GP6A ALH+	30ugF1-WV	1,310,720	20,480
12B	7699	GP6A ALH+	30ugF1-WV	655,380	20,480
13A	7700	GP6A ALH+	30ugF1-WV	1,310,720	5,120
13B	7701	GP6B ALH+	30ugF1-WV	1,310,720	20,480
14A	7702	GP6B ALH+	30ugF1-WV	1,310,720	20,480
14B	7703	GP6B ALH+	30ugF1-WV	1,310,720	2,560
15A	7704	GP6B ALH+	30ugF1-WV	1,310,720	10,240
15B	7705	GP6B ALH+	30ugF1-WV	1,310,720	2,560
16A	7706	GP7 Plaque USP		320	735
16B	7707	GP7 Plaque USP		1,280	320
17A	7708	GP7 Plaque USP		640	10,240
17B	7709	GP7 Plaque USP		640	20,480
18A	7710	GP7 Plaque USP		1,280	1,280
18B	7711	GP7 Plaque USP		640	1,280
19A	7712	GP7 Plaque USP		640	5,120
19B	7713	GP7 Plaque USP		320	2,560
20A	7714	GP7 Plaque USP		1,280	5,120
20B	7715	GP7 Plaque USP		1,280	40,960
21A	7716	GP8 Alhydrogel		640	320
21B	7717	GP8 Alhydrogel		640	320
22A	7718	GP8 Alhydrogel		320	320
22B	7719	GP8 Alhydrogel		640	320
23A	7720	GP8 Alhydrogel		640	320
23B	7721	GP8 Alhydrogel		320	320
24A	7722	GP8 Alhydrogel		320	320
24B	7723	GP8 Alhydrogel		1,280	320
25A	7724	GP8 Alhydrogel		320	320
25B	7725	GP8 Alhydrogel		320	320
33B	F/V POOL (+ CONTROL)			655,380	81,920

Mouse weights for aerosol run of 23Feb96

24.9
30.9 average wt (gm)
29.98
32.5 Cage 9A
32.4
29.5
28.8
28.2
30.1
34.3
28.2

Ig Subclass ELISA response to vaccination with *Yersinia pestis* V and F1 antigens (continued)

Plate setup:

Same as 21 FEB 96, except test samples (9) for V antigen only.

Procedure:

Same as 21.FEB 96, except:

•Second antibodies (goat anti-mouse IgG1, IgG2a, and IgG2b) dilution 1:3000 (decreased from 1:2000).

•Conjugate dilution decreased to 1:3000 (from 1:2000)...

Results: see separate pages.

See page 137 and back 3175

BALB/c day 58 postimmunization

Sample	Treatment	IgG1(H+L)	IgG1	IgG2a	IgG2b
2041	Alhydrogel	320	320	320	640
2042	Alhydrogel	320	320	320	320
2043	Alhydrogel	320	320	320	320
2044	Alhydrogel	320	320	320	320
2045	Alhydrogel	320	320	320	320
2046	Alhydrogel	320	320	320	320
2047	Alhydrogel	320	320	320	320
2048	Alhydrogel	320	320	320	320
2049	Alhydrogel	320	320	320	320
2051	Alh+V	640,000	655,380	20,480	31,920
2062	Alh+V	320,000	1,310,720	81,920	10,960
2063	Alh+V	640,000	1,310,720	40,960	31,920
2064	Alh+V	320,000	655,380	40,960	40,960
2065	Alh+V	640,000	327,680	31,920	20,480
2067	Alh+V	1,280,000	1,310,720	31,920	31,920
2058	Alh+V	1,280,000	1,310,720	40,960	16,0160
2069	Alh+V	640,000	1,310,720	20,480	31,920

On return, all
samples b640

AERIAL EXPOSURE SHEET

Antibiotic Treatment / Vaccine Challenge

Date: 23-Feb

PI: COL Byrne / LTC Anderson

Agent: Plague

Strain: CO92

Animal Model: Mouse

Strain: Swiss Webster

Wt: (Ave.) : 20g / 29.98g

CO92 LD50=2.1E+04

Sex: female

Run #	cfu/l				Inhaled Dose		
	AGI/ml	AGI	aerosol	MV	cfu	LD50s	
1	3.50E+07	3.50E+08	5.83E+06	0.02	1.17E+06	55.56	CO92
2	3.80E+07	3.80E+08	6.33E+06	0.02	1.27E+06	60.32	CO92
3	5.10E+07	5.10E+08	8.50E+06	0.02	1.70E+06	80.95	CO92
4	4.00E+07	4.00E+08	6.67E+06	0.02	1.33E+06	63.49	CO92
5	8.10E+06	8.10E+07	1.35E+06	0.02	2.70E+05	12.86	CO92
6	1.70E+07	1.70E+08	2.83E+06	0.027	7.65E+05	36.43	CO92
7	1.60E+07	1.60E+08	2.67E+06	0.027	7.20E+05	34.29	CO92

Date: 23Feb96		Project: Active F1-WV, F1+ Long-term study Day 449 Postimmunization			
Notebook #: 3739					
Inoculum: Yersinia pestis strain, CO92					
Route: aerosol	Dose: as shown below	LD ₅₀ , 34-36			
Animal strain: Swiss Webster	Arrival 10/17/95 at 7-8wks		Vendor: Harlan Sprague Dawley	Sex: female	
Month - Feb Day -	23 24 25 26 27 28 29 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23				
Day postinfection	0 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29				Comments/Chip #
Group	LD50				
9A 10ugF1+ 20ug mauro-V	CO92	WHITE	BLACK		221D54560C/LT-081 2221523113A/LT-082 221D4E207A/LT-083 2225395749/LT-084 2221340347/LT-085 2221774E53/LT-086 2222576612/LT-087 22224E6F58/LT-088 221D563E57/LT-089 22212D0F34/LT-090 221D681542/LT-221 22280E3128/LT-222 221D584D78/LT-223 221D4C4067/LT-224 22212A2966/LT-225
9B 10ugF1+ 20ug mauro-V	CO92	RED	BLACK		
10A 30ugF1- WV	CO92	RED	BLACK		221D62392D/LT-091 2222411F74/LT-092 221D4D6C2C/LT-093 221D554048/LT-094 2221473A36/LT-095 2221240C60/LT-096 22214F1224/LT-097 2221425212/LT-098 22223C390C/LT-099 2221414605/LT-100 222124374C/LT-226 2222586303/LT-227 2222371808/LT-228 2222443D5B/LT-229 22213B707C/LT-230
10B 30ugF1- WV	CO92	WHITE	RED		
11 Plague USP Greer 1128X1	CO92	WHITE	BLACK		221D6F12X01/LT-101 2221305178/LT-102 2221720E09/LT-103 221D403C64/LT-104 2222355935/LT-105 221D4003064/LT-106 2221633E37/LT-107 22254A6B01/LT-108 2221475264/LT-109 2221760056/LT-110 2222401108/LT-111 2222422877/LT-112 2227610A1G/LT-113 lost 221D686900/LT-114 221D703E2B/LT-115 221D871449A/LT-116 22225C0824/LT-117 2221586D78/LT-118 22214C97104/LT-119 221D753C07/LT-120
12 Alhydrogel alone	CO92	BLACK	BLACK		
13 MTPF10 7.5E-1.9 1128X1	CO92	GREEN	BLACK		
14 MTPF10 7.1E-1 1128X1	CO92	GREEN	BLACK		

For Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

Discard dead animals

Check chip number of dead mice with scanner

Mark number of animals alive in each cage

LOST after Chip # means the chip has fallen out

HIGH COLOR
BLACK - AEROSOL RUN 6
RED - AEROSOL RUN 7

SGT LIVINGSTON L VITALE

A pencil mark, spanning my line of authorship, happened to run across me

Exhibit GA15

ELISA V Summary *See page 75-8102* 3 APR 96

PROTOCOL: V LONGTERM				V antigen TITER	
Plate	Serum	Group	Treatment	Bleed	IgG(H+L)
1A	8288	GP9A	F1+V	+DAY86	81,920
1B	8289	GP9A	F1+V	+DAY86	327,680
2A	8290	GP9A	F1+V	+DAY86	40,960
2B	8291	GP9A	F1+V	+DAY86	NO SERUM
3A	8292	GP9A	F1+V	+DAY86	163,840
3B	8293	GP9A	F1+V	+DAY86	327,680
4A	8294	GP9A	F1+V	+DAY86	327,680
4B	8295	GP9A	F1+V	+DAY86	655,360
5A	8296	GP9A	F1+V	+DAY86	40,960
5B	8297	GP9A	F1+V	+DAY86	655,360
6A	8298	GP9B	F1+V	+DAY86	327,680
6B	8299	GP9B	F1+V	+DAY86	327,680
7A	8300	GP9B	F1+V	+DAY86	81,920
7B	8301	GP9B	F1+V	+DAY86	655,360
8A	8302	GP9B	F1+V	+DAY86	327,680
Geomean					220,512
8B	8303	GP10A	F1-WV	+DAY86	163,840
9A	8304	GP10A	F1-WV	+DAY86	163,840
9B	8305	GP10A	F1-WV	+DAY86	163,840
10A	8306	GP10A	F1-WV	+DAY86	163,840
10B	8307	GP10A	F1-WV	+DAY86	163,840
11A	8308	GP10A	F1-WV	+DAY86	81,920
11B	8309	GP10A	F1-WV	+DAY86	163,840
12A	8310	GP10A	F1-WV	+DAY86	163,840
12B	8311	GP10A	F1-WV	+DAY86	163,840
13A	8312	GP10A	F1-WV	+DAY86	163,840
13B	8313	GP10B	F1-WV	+DAY86	81,920
14A	8314	GP10B	F1-WV	+DAY86	327,680
14B	8315	GP10B	F1-WV	+DAY86	327,680
15A	8316	GP10B	F1-WV	+DAY86	163,840
15B	8317	GP10B	F1-WV	+DAY86	327,680
Geomean					171,589
16A	8318	GP11	PLAQUE USP	+DAY86	1,280
16B	8319	GP11	PLAQUE USP	+DAY86	640
17A	8320	GP11	PLAQUE USP	+DAY86	1,280
17B	8321	GP11	PLAQUE USP	+DAY86	640
18A	8322	GP11	PLAQUE USP	+DAY86	1,280
18B	8323	GP11	PLAQUE USP	+DAY86	1,280
19A	8324	GP11	PLAQUE USP	+DAY86	1,280
19B	8325	GP11	PLAQUE USP	+DAY86	640
20A	8326	GP11	PLAQUE USP	+DAY86	640
20B	8327	GP11	PLAQUE USP	+DAY86	1,280
Geomean					970
21A	8328	GP12	ALHYDRO ALONE	+DAY86	640
21B	8329	GP12	ALHYDRO ALONE	+DAY86	640
22A	8331	GP12	ALHYDRO ALONE	+DAY86	640
22B	8333	GP12	ALHYDRO ALONE	+DAY86	1,280
23A	8335	GP12	ALHYDRO ALONE	+DAY86	1,280
23B	8336	GP12	ALHYDRO ALONE	+DAY86	1,280
24A	8337	GP12	ALHYDRO ALONE	+DAY86	640
Geomean					861
24B	F1/V POOL				655,360

500ml, undiluted

9AFR 96

| Hbt |
|-------|-------|-------|-------|-------|-------|-------|-------|-------|--------|
| No: 1 | No: 2 | No: 3 | No: 4 | No: 5 | No: 6 | No: 7 | No: 8 | No: 9 | No: 10 |
| 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 | 1 |
| a | a | a | a | a | a | a | a | a | a |
| 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 | 2 |
| b | b | b | b | b | b | b | b | b | b |
| 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 | 3 |
| A | A | A | A | A | A | A | A | A | A |
| E | E | E | E | E | E | E | E | E | E |
| M | M | M | M | M | M | M | M | M | M |
| K | K | K | K | K | K | K | K | K | K |
| λ | λ | λ | λ | λ | λ | λ | λ | λ | λ |

MAb V-YP 7F5-1-1, SUBCLONES -10

ELISA F1 Summary

3 APR 96

PROTOCOL: V/F1 LONGTERM				F1 antigen TITER	
Plate	Serum	Group	Treatment	Bleed	IgG(H+L)
1A	8288	GP9A	F1+V	+DAY86	NSUFFICIENT SERUM
1B	8289	GP9A	F1+V	+DAY86	20,480
2A	8290	GP9A	F1+V	+DAY86	NSUFFICIENT SERUM
2B	8291	GP9A	F1+V	+DAY86	NO SERUM
3A	8292	GP9A	F1+V	+DAY86	40,960
3B	8293	GP9A	F1+V	+DAY86	2,560
4A	8294	GP9A	F1+V	+DAY86	20,480
4B	8295	GP9A	F1+V	+DAY86	5,120
5A	8296	GP9A	F1+V	+DAY86	20,480
5B	8297	GP9A	F1+V	+DAY86	81,920
6A	8298	GP9B	F1+V	+DAY86	40,960
6B	8299	GP9B	F1+V	+DAY86	2,560
7A	8300	GP9B	F1+V	+DAY86	20,480
7B	8301	GP9B	F1+V	+DAY86	5,120
8A	8302	GP9B	F1+V	+DAY86	5,120
Geomean					15,343
8B	8303	GP10A	F1-WV	+DAY86	5,120
8B	8304	GP10A	F1-WV	+DAY86	20,480
9A	8305	GP10A	F1-WV	+DAY86	1,280
9B	8306	GP10A	F1-WV	+DAY86	10,240
10A	8307	GP10A	F1-WV	+DAY86	10,240
10B	8308	GP10A	F1-WV	+DAY86	2,560
11A	8309	GP10A	F1-WV	+DAY86	2,560
11B	8310	GP10A	F1-WV	+DAY86	5,120
12A	8311	GP10A	F1-WV	+DAY86	1,280
12B	8312	GP10A	F1-WV	+DAY86	5,120
13A	8313	GP10B	F1-WV	+DAY86	1,280
13B	8314	GP10B	F1-WV	+DAY86	2,560
14A	8315	GP10B	F1-WV	+DAY86	5,120
14B	8316	GP10B	F1-WV	+DAY86	5,120
15A	8317	GP10B	F1-WV	+DAY86	10,240
Geomean					4,256
15B	8318	GP11	PLAQUE USP	+DAY86	5,120
16A	8319	GP11	PLAQUE USP	+DAY86	5,120
16B	8320	GP11	PLAQUE USP	+DAY86	10,240
17A	8321	GP11	PLAQUE USP	+DAY86	10,240
17B	8322	GP11	PLAQUE USP	+DAY86	10,240
18A	8323	GP11	PLAQUE USP	+DAY86	5,120
18B	8324	GP11	PLAQUE USP	+DAY86	10,240
19A	8325	GP11	PLAQUE USP	+DAY86	20,480
19B	8326	GP11	PLAQUE USP	+DAY86	5,120
20A	8327	GP11	PLAQUE USP	+DAY86	10,240
Geomean					8,317
20B	8328	GP12	ALHYDRO ALONE	+DAY86	320
21A	8329	GP12	ALHYDRO ALONE	+DAY86	320
21B	8331	GP12	ALHYDRO ALONE	+DAY86	320
22A	8333	GP12	ALHYDRO ALONE	+DAY86	320
22B	8335	GP12	ALHYDRO ALONE	+DAY86	320
23A	8336	GP12	ALHYDRO ALONE	+DAY86	320
23B	8337	GP12	ALHYDRO ALONE	+DAY86	320
Geomean					320
24A	F1/V POOL				40,960

On new paper
all 640 new
96

Exhibit GA16

		Mean time-to-death (MTD)															
Challenge	group	1	2	3	4	5	6	7	8	9	10	MTD	+/- Std	Std	Group		
Subcutaneous:																	
1	alhydrogel alone, days 0, 30, sc																
C12	100	5	6	6	7	7	8	8	8	10	11	7.6	1.837873	0.61262	1		
2	alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12	100	28	28	28	28	28	28	28	28	28	28	28	25.8	6.957011	2.319	3	
3	alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc																
C12	Max	6	28	28	28	28	28	28	28	28	28	28	28	25.8	6.957011	2.319	4
4	alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12	Max	6	28	28	28	28	28	28	28	28	28	28	28	25.8	6.957011	2.319	4
5	alhydrogel + 27.2 µg F1-WV fusion protein day 0, 30, sc																
C12	Max	28	28	28	28	28	28	28	28	28	28	28	28	0	0	5	
6	alhydrogel alone, days 0, 30, sc																
C12	Max	3	3	3	3	3	3	3	3	3	5	5	5	3.6	0.966092	0.32203	6
7	alhydrogel + 27.2 µg F1-WV fusion protein day 0, 30, sc																
CO92	100	28	28	28	28	28	28	28	28	28	28	28	28	0	0	7	
8	alhydrogel alone, days 0, 30, sc																
CO92	100	3	3	4	4	4	4	6	6	6	7	7	4.8	1.549193	0.5164	8	
Aerosol:																	
9	alhydrogel alone, days 0, 30, sc																
C12	50	3	3	3	3	3	3	3	3	3	3	3	0	0	0	9	
10	alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12	50	28	28	28	28	28	28	28	28	28	28	28	0			10	
11	alhydrogel + 10 µg Mauro-V urea, days 0, 30, sc																
C12	Max	5	28	7	28	28	28	28	28	28	28	28	23.6	9.287985	3.096	11	
12	alhydrogel + 13.6 µg F1-WV fusion protein day 0, 30, sc																
C12	Max	28	28	28	28	28	28	28	28	28	28	28	0	0	0	12	

Challenge	1	2	3	4	5	6	7	8	9	10	MTD	MTD	Std
group											+/- Stddev		error
13 alhydrogel + 27.2 μ g F1-WV fusion protein day 0, 30, sc													
C12 Max	28	28	28	28	28	28	28	28	28	28	28	0	0
14 alhydrogel alone, days 0, 30, sc													
C12 Max	3	3	3	3	3	3	3	3	3	4	3.11111	0.333333	0.11785
15 alhydrogel + 13.6 μ g F1-WV fusion protein day 0, 30, sc													
CO92 100	28	28	28	28	28	28	28	28	28	28	28	0	0
16 alhydrogel alone, days 0, 30, sc													
CO92 100	28	4	3	3	3	3	3	3	3	3	5.6	7.87683	2.62561
17 alhydrogel + 10 μ g F1 + 10 μ g Mauro's V, days 0, 30, sc													
C12 Max	28	28	28	28	28	28	28	28	28	28	28	0	0
18 Greer plague vaccine, days 0, 30, sc													
C12 Max	4	4	3	3	3	3	3	3	3	3	3.25	0.46291	0.17496
19 alhydrogel alone, days 0, 30, sc													
C12 Max	3	3	4	4	4	5					3.8	0.83666	0.41833
													19

Date: 28Jun96	Project: Longterm F1-V, F1+V Challenge, Day 216		
Notebook #: 3739		Inoculum: Yersinia pestis strain, CO92	
Route: aerosol		Dose: as shown below	
Animal strain: Swiss Webster	Arrival 10/17/95 at 7-8wks	Vendor: Harlan Sprague Dawley	Sex: female
Month - Jun Day	28 29 30 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29		
Day postinfection	0 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29		Comments/Chip #
Group	LD50		
GP13A	CO92	WHITE	2227712235/LT-121
10ug F1 + 50 ug V hydrogel			222241257BL/LT-122
			22224F1410/LT-123
			221D744B1A/LT-124
			22224E7477/LT-125
			22224C4154/LT-126
			221D66020D/LT-127
			2221427332/LT-128
			221D730F06/LT-129
			221D691F24/LT-130
GP13B	CO92	WHITE	221D737744/LT-231
10ug F1 + 50 ug V hydrogel			2221313035/LT-232
			2222392702/LT-233
			221D73716A/LT-234
			221D527D35/LT-235
GP14A	CO92	RED	2216B396D/LT-131
10ug F1 + 50 ug V hydrogel			221B37023F/LT-132
			222258140A/LT-133
			22222E6823/LT-134
			2221431C17/LT-135
			221D757C7E/LT-136
			2222582B00/LT-137
			2221754939/LT-138
			221D5B0C51/LT-139
			222153781B/LT-140
GP14B	CO92	BLUE	221B2E7B71/LT-236
10ug F1 + 50 ug V hydrogel			221D484645/LT-237
			22276D3C14/LT-238
			221D4C655D/LT-239
			2222516E3F/LT-240
GP15B	CO92	BROWN	221D481476/LT-141
10ug F1 + 50 ug V hydrogel			221D69771B/LT-142
			2228169110/LT-143
			2222526107/LT-144
			22217D4C57/LT-145
			22222E5018/LT-146
			221B2C4420/LT-147
			222156603B/LT-148
			2221522454/LT-149
			222168412F/LT-150
			2221477A4F/LT-151
			222258113S/LT-152
			2222394F69A/LT-153
			2221261D5A/LT-154
			2221793120/LT-155
			221D725567/LT-156
			221D5E4642/LT-157
			222126640B/LT-158
			22274C6575/LT-159
			22252D2519/LT-160

Animal Caretakers: Contact LTC Anderson, Office Ext 4933, Fax Ext 2152, Home (301) 473-5059 if you have any questions

dead animals

chip number of dead mice with scanner

number of animals alive in each cage

Chip # means the chip has fallen out

HEAD COLOR
WHITE Run 1 AF-RUGOL
BLACK Run 2 AF-RUGOL

grey

black

Mouse weight for several challenges of 28 Jan 51
34.2, 32.9, 25.7, 35.5, 40.0, 39.0, 27.4, 31.7, 25.2, 41.0 gm = 34.7 avg gm

The counts were as follows:

Prespray -- 4.2×10^9 (48/34/43 on 10e7 plate)

AGE's
#1 -- 2.6×10^7 (18/36/25 on 10e5 plate)

#2 -- 2.2 x 10e7 (222/196/229 on 10e4 plate)

fuann

AFG: 11 EXPOSURE SHEET

Animal exposure #: 96-0301f	Location: A-4	Date: 26-Jun-96									
Approval Operator: SPC Holmes		Agent: Plaza - COSA									
Operational Check Performed: ✓		Protocol #:									
Exposure System Type#: Non-Dry		P.I.: LTC Anderson									
System Flow Rate: 1A Lpm	Timer Check: ✓ Pre ✓ Post										
Collision #: 5	Start time 0910	1104									
Panel #: 3	End time 0916	1109									
Electronic Flow Meter #: E0979	OK										
Run #	Animal Species	Start T			5 min T			Start Time	AGI #	Comments	
		Dry	Wet	Rel. Hum.	Dry	Wet	Rel. Hum.				
1	24	mic	73	68	77%	74	71	85%	0945	366	1 mouse Dead on arrival
2	25	mic	73	68	77%	74	71	85%	1031	213	1 Dead mouse during aerosol
2 - 10. 6/25											
25 Jun 96											

Active Immunization

Date: 28-Jun

PI: LTC Anderson
Agent: Plague
Strain: CO92

Animal Model: Mouse

Wt: (Ave.) 34.7g

CO92 LD50 = 2.1E + 04

Run #	AGI	AGI	Aerosol	IV	Inhaled Dose	Plague	
	cfu / ml	cfu	cfu / l	l	cfu	LD50s	Strain
1	2.60E+07	2.60E+08	4.33E+06	0.03	1.30E+06	61.90	CO92
2	2.20E+07	2.20E+08	3.67E+06	0.03	1.10E+06	52.38	CO92

Plate	Bleed: Serum	Group	Day: Day +204	26JUN96	25JUN96
				TREATMENT	F1 TITER
1A	8906	GP13A	10F1+20V	10,240	20,480
1B	8907	GP13A	10F1+20V	1,280	40,960
2A	8908	GP13A	10F1+20V	10,240	81,920
2B	8909	GP13A	10F1+20V	5,120	20,480
3A	8910	GP13A	10F1+20V	20,480	20,480
3B	8911	GP13A	10F1+20V	10,240	40,960
4A	8912	GP13A	10F1+20V	40,960	10,240
4B	8913	GP13A	10F1+20V	2,560	81,920
5A	8914	GP13A	10F1+20V	10,240	81,920
5B	8915	GP13A	10F1+20V	5,120	20,480
6A	8916	GP13B	10F1+20V	20,480	81,920
6B	8917	GP13B	10F1+20V	2,560	40,960
7A	8918	GP13B	10F1+20V	10,240	163,840
7B	8919	GP13B	10F1+20V	20,480	81,920
8A	8920	GP13B	10F1+20V	10,240	81,920
8B	8921	GP14A	30ugF1-V	2,560	81,920
9A	8922	GP14A	30ugF1-V	2,560	81,920
9B	8923	GP14A	30ugF1-V	1,280	40,960
10A	8924	GP14A	30ugF1-V	1,280	40,960
10B	8925	GP14A	30ugF1-V	1,280	81,920
11A	8926	GP14A	30ugF1-V	2,560	81,920
11B	8927	GP14A	30ugF1-V	5,120	81,920
12A	8928	GP14A	30ugF1-V	1,280	40,960
12B	8929	GP14A	30ugF1-V	2,560	163,840
13A	8930	GP14A	30ugF1-V	5,120	81,920
14A	8931	GP14B	30ugF1-V	10,240	163,840
14B	8932	GP14B	30ugF1-V	20,480	81,920
15A	8933	GP14B	30ugF1-V	10,240	655,360
15B	8934	GP14B	30ugF1-V	5,120	81,920
16A	8935	GP14B	30ugF1-V	2,560	40,960
16B	8936	GP15	PlagueUSP	2,560	2,560
17A	8937	GP15	PlagueUSP	2,560	2,560
18A	8938	GP15	PlagueUSP	2,560	2,560
18B	8939	GP15	PlagueUSP	5,120	2,560
19A	8940	GP15	PlagueUSP	2,560	2,560
19B	8941	GP15	PlagueUSP	5,120	2,560
20A	8942	GP15	PlagueUSP	5,120	2,560
20B	8943	GP15	PlagueUSP	20,480	5,120
21A	8944	GP15	PlagueUSP	40,960	640
21B	8945	GP15	PlagueUSP	5,120	1,280
22A	8946	GP16	ALH alone	640	2,560
22B	8947	GP16	ALH alone	320	640
23A	8948	GP16	ALH alone	320	640
23B	8949	GP16	ALH alone	320	1,280
24A	8950	GP16	ALH alone	320	640
24B	8951	GP16	ALH alone	640	640
25A	8952	GP16	ALH alone	320	1,280
25B	8953	GP16	ALH alone	1,280	2,560
26A	8954	GP16	ALH alone	640	640
26B	8955	GP16	ALH alone	1,280	5,120
27A	F1/V	POOL		327,680	1,310,720
27B	Norm Mouse	POOL		120	2,560

GEOMEAN:	Group	TREATMENT	F1 TITER	V TITER
	GP13A/B	10F1+20V	8,12	44,926
	GP14A/B	30ugF1-V	3,378	85,794
	GP15	PlagueUSP	5,37	2,229
	GP 16	ALH alone	120	1,194

For plaque titration of 5 of 76
 140 070 0092
 74 615 012

data from Dr. Walker
 7/26
 8/18/96

Steyer
plague-challenge.sc 7/5/96

7/5/96

T

P.I. = LTC George Anderson
40 mice, C092 - 100 LD50s
30 mice, C12 - 100 LD50s

Parenteral challenge of mice with C092/M.S. and C12/M.S.

1. Streak 1 slant each with the Master Seed of C092 and C12.
Incubate 2 days at room temperature.
2. Harvest by suspending in 4-5 mls of HIB.
3. Read OD620 of a 1/10 dilution.
4. Adjust to OD 1.0

7/5/96:

Adjusted ODs and read final ODs on 1/2 dilutions:

Final OD = 1.064, for C092

" " - 1.10, for C12

C092/M.S.:

1. Prepare dose

5.0 - 7.5x10e2/ml:

- (1) Add 0.2 ml OD 1.0 to 1.8 mls HIB.
- (2) Add 0.2 ml (1) to 1.8 mls HIB.
- (3) Add 0.5 ml of (2) to 4.5 mls HIB.
- (4) Add 0.5 ml of (3) to 4.5 mls HIB.
- (5) Add 0.5 ml of (4) to 4.5 mls HIB.
- (6) Add 4.0 ml of (5) to 36 mls HIB - - Pipet 10 mls into each of 3 tubes:

mice INOCULUM: $1 \times 10e3/ml$: ~200 cfu/dose

2. Plating: The sample will be diluted in HIB and plated on SBAP:

Total No.

<u>suspension</u>	<u>Conc./ID</u>	<u>dilution</u>	<u>no. plates</u>	<u>plates</u>
mice Inoculum	$5 \times 10e2/ml$	undil, 10-1	5 each	10

RESULTS:

7/5/96 doses: 1.4 x 10e3/ml, 280 cfu/dose (140 LD50s)

7/12/96 doses: 6.5 x 10e2/ml, 130 cfu/dose (72 LD50s)

7/18/96 doses: x 10e2/ml, cfu/dose (LD50s) - *correlated*

C12/M.S.:

1. Adjust slant suspension to OD620 = 1.0.

Prepare dose

2.3 x 10e3/ml:

Moore
34, 2

The count
Pres
AGC
#1
#2

- (1) Add 0.2 ml OD 1.0 to 1.8 mls HIB.
- (2) Add 0.2 ml (1) to 1.8 mls HIB.
- (3) Add 0.5 ml of (2) to 4.5 mls HIB.
- (4) Add 0.5 ml of (3) to 4.5 mls HIB.
- (5) Add 1.0 ml of (4) to 9.0 mls HIB.
- (6) Add 6.0 ml of (5) to 18 mls HIB (1/4) -

INOCULUM, C12-100 sc LD₅₀s (910 cfu). Pipet 10
mls into each of 2 tubes:
1 x 10e3/ml: ~200 cfu/dose

2. Plating: The Inoculum will be diluted in HIB and plated on SBAP:

<u>suspension</u>	<u>Conc./ID</u>	<u>dilution</u>	<u>Total No.</u>	<u>no. plates</u>	<u>plates</u>
C12 Inoculum	2.3x10e3/ml	undil.		5	
		10-1		5	
		10-2		5	
					TOTAL-15

RESULTS:

7/5/96 doses:

3.36 x 10e3/ml. 6.7 x 10e2 cfu/dose (73.6 LD₅₀s)

7/12/96 doses:

3.0 x 10e3/ml. 5.8 x 10e2 cfu/dose (63.7 LD₅₀s)

7/18/96 doses:

x 10e3/ml. x 10e2 cfu/dose (LD₅₀s) - *consider*

MAR-12-1996 15:45 FROM WADSWORTH DAI

TO 88723167037581222 P.02

FROM : AIBS RESTON

PHONE NO. : 703 758 1222

Mar. 11 1996 04:38PM P2

Exhibit GA19

**AIBS PEER REVIEW TO USAMRMC
MEDICAL BIOLOGICAL DEFENSE RESEARCH PROGRAM
ON PLAGUE**

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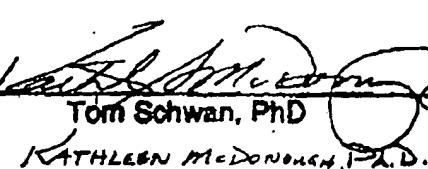
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APPROVED:


Tom Schwan, PhD
KATHLEEN MCDONOUGH, PhD

DATE: March 12, 1996

INTRODUCTION

AIBS was requested by US Army Medical Research and Development Command (USAMRDC) to convene a review Panel to provide an assessment of the scientific merit of the Medical Biological Defense Research Program (MBDRP) on Plague. It was requested that the three scientific reviewers have a collective knowledge of the following subject areas: *Yersinia pestis*, Vaccine Production, Molecular Genetics and FDA requirements for a vaccine. Such a panel was convened and provided with documentation by USAMRDC to read prior to the conference. This consisted of abstracts prepared by the individual investigators who form the MBDRP on Plague (see Appendix 1.)

CHARGE TO PANEL

Three scientific reviewers were asked to evaluate the MBDRP on Plague. They independently reviewed material provided by USAMRDC and attended a conference on the subject matter. They were asked to judge the scientific merits of the Program.

The reviewers, individually, provided comments to AIBS, who in turn compiled this written report summarizing these comments and the discussions at the conference. The Chairman of the Review Panel read and approved the report prior to its submission to USAMRDC.

PRESENTATION SUMMARIES

The conference comprised presentations by each of the following investigators. Abstracts were provided for by each and are attached as Appendix 1.

COL ARTHUR FRIEDLANDER
Overview of plague program

COL RUSSELL BYRNE
Antibiotic treatment of experimental pneumonic plague

DR. PATRICIA WORSHAM, DR. M. LOUISE PITT, LTC KELLY DAVIS
F1 is not a required virulence factor for the mouse or non-human primate

MAJ GERALD P. ANDREWS, LTC GEORGE J. ANDERSON, JR.
Protective efficacy of active immunization with purified F1 from *Yersinia pestis* and an *Escherichia coli* recombinant strain against lethal parenteral and respiratory plague challenge

DR. PATRICIA WORSHAM
Studies on the role of the pigmentation locus in the pathogenesis of *Y. pestis*

DR. SUSAN L. WELKOS, LTC KELLY J. DAVIS
Analysis of the role of pPst encoded genes in pathogenesis of infection by *Y. pestis*

DR. ALAN SAMPLE

Plasminogen activator protease degrades proinflammatory cytokines

MAJ GERALD P. ANDREWS, DR. SUSAN STRALEY, DR. ALAN SAMPLE, MAJ GERALD P. ANDREWS

Cloning, Expression, Purification, and Protective Efficacy of Yops and pH 6 antigen

LTC GEORGE J. ANDERSON, JR., DR. DAVID HEATH

Cloning, expression, and protective efficacy of V antigen

LTC GEORGE J. ANDERSON, JR.

Cloning, expression, and protective efficacy of F1-V fusion protein

DR. JEFFREY PULLEN

Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Yersinia pestis*

COL ARTHUR FRIEDLANDER

Overview of future plans

SUMMARY EVALUATIONS OF THE RESEARCH AREAS

The review panel read the abstracts provided by the investigators prior to the meeting on February 15, 1996, and listened to presentations by each of the investigators at the meeting. The following comments include recommendations to individual investigators, and are intended to be constructive. Certain points apply to more than one project, or even to the program as a whole, and hence may appear repetitive. Also, the reviewers recognize that some of their recommendations may be affected by programmatic decisions that are beyond the control of the immediate Program staff and thus may not prove to be possible.

COL ARTHUR FRIEDLANDER

Overview of plague program

The USAMRMC Plague Research Program's primary objective is to develop a vaccine that will protect military personnel if exposed to an aerosol attack of *Yersinia pestis*, the causative agent of plague. Given that the currently available vaccine (USP) protects primarily through anti-F1 antibody, that this vaccine offers very poor protection from primary pneumonic plague, and that F1⁻ strains are highly virulent, there clearly is a need for a new, more protective vaccine. Once developed, the general population living in areas endemic for plague would also benefit from such a vaccine.

Most of the projects presented as separate studies and presentations clearly meet the program's primary objective. Part of the rationale for the approach taken is that an aerosol attack of *Yersinia pestis* might include strains that do not produce the F1 capsular antigen. Given that the current vaccine (USP) stimulates primarily antibodies

**AIBS PEER REVIEW TO USAMRDC
MEDICAL BIOLOGICAL DEFENSE RESEARCH PROGRAM
ON PLAGUE**

TIME: February 15, 1996, 8.00am to 5.00 pm

LOCATION: US Army Medical Research Institute of Infectious Diseases,
Conference Room, Fort Detrick, Frederick, MD

EXECUTIVE SUMMARY

Overall, the program has made very significant and impressive advances in only a few years towards the development of a new vaccine, and Dr. Friedlander and his entire team of investigators can be proud of their accomplishments to date. They clearly have a very viable, sound program with a good team of investigators that is focused with high potential to succeed. It is hoped that the administration will continue to support this effort and provide the group with the resources and time necessary to complete their task. The investigators clearly considered the recommendations of the previous reviewers and incorporated several of the suggestions into their program.

The team has invested significant effort in examining numerous virulence determinants of *Yersinia pestis* for their ability to stimulate protection through immunization. The F1 capsular antigen and the V antigen have been shown by investigators in other laboratories to be good candidates for inclusion in a new multivalent subunit vaccine. The team at USAMRMC has confirmed the protective value of these two antigens. However, realizing that F1 and V antigens might not be sufficient for full protection against all virulent strains of *Y. pestis*, the group has worked through an impressive list of additional candidates. The only other antigen that offered significant protection was YopD, although protection was only observed when mice were challenged with the F1⁻ strain. Passive immunizations with anti-F1 and anti-YopM antisera deserve further attention. Combined antibiotic treatment and immunization might increase the survival of animals challenged by aerosol.

The team appears to make use of mice and nonhuman primates as excellent animal models for both their parenteral and aerosol challenge experiments. The current vaccine study protocols for test challenges are very good.

The development of *in vitro* correlates of immunity should be a high priority of the program. It is currently the weakest portion of the future plans. As discussed with the investigators, the assumption that protection is solely antibody-mediated has potential difficulties. Before continuing studies to map active B cell epitopes, the investigators need to determine the role of T cells in immunity to plague.

examined for *Y. pestis* in the LD50 studies and survivors were examined for clearance of the organisms to determine the full level of protection provided by vaccination.

In the first study, the V antigen was examined for its ability to generate a protective immune response in mice challenged by parenteral subcutaneous or aerosol challenge with either the F1⁺ or F1⁻ isogenic strains of *Y. pestis*. Recombinant V antigen was cloned and expressed in two fusion/expression systems and used with an adjuvant approved for human use (Alhydrogel). Both preparations of rV antigen were administered twice and provided very good protection in mice challenged by both routes and both strains. This is an excellent study and identifies (as another laboratory has demonstrated independently) the V antigen as an excellent candidate immunogen to include in a vaccine to protect from aerosol infections with either F1⁺ or F1⁻ strains. These studies are critical to the program's objective and provides some quite exciting results.

The second study extends the work on the V antigen of *Y. pestis* by examining protection following a single dose of 10 µg (the previous study used two immunizations prior to challenge). Mice were subsequently challenged by aerosol exposure to either low and high doses of the F1⁺ or F1⁻ strain. Protection ranged from 70% to 78% survival in these mice, demonstrating that a single immunization could afford significant protection from an aerosol route of infection. However, the schedule including a primary immunization followed by a single boost afforded 20% to 30% greater protection (previous report). While it is of interest what level of protection results from a single dose, future work with nonhuman primates will likely confirm what we know about many other bacterial vaccines, i.e., better protection results with boosts following the primary immunization.

Two areas need to be addressed in future work on the V antigen. The studies presented used the V antigen tagged with histidine from the pET vector. If this antigen is to be used in humans, a method for the efficient removal of the his-tag is needed. Identifying the active sites on the V antigen responsible for protective immunity as well as potential negative biological activities, such as immune suppression, may be required for this antigen to be safe. The group might also consider examining how long protective immunity lasts following vaccination with the V antigen. Some of these issues were addressed by Dr. Friedlander in his closing remarks.

LTC GEORGE J. ANDERSON, JR.

Cloning, expression, and protective efficacy of F1-V fusion protein (abstract 17)

Prior studies have confirmed the potential for both F1 and V antigen to protect mice from *Y. pestis* by both parenteral and aerosol routes. In this study a construct was made containing the F1 and V antigen genes for expression of a fusion protein. When the F1-V fusion protein was used for immunization, mice were protected when challenged by needle or aerosol with either the F1 positive or F1 negative strain of *Y. pestis*. Poorer protection resulted when only a portion of the V antigen was expressed as a fusion protein with F1. This work is quite clever and interesting, and advances the program's effort towards the development of a multivalent vaccine. The attempt to make fusions of these two antigens also demonstrates an advance towards reducing

the steps required for making and purifying antigens for the vaccine. The investigators are also testing longer term antibody responses and how long protection lasts (a concern raised from the previous studies with the V antigen alone). Antibody responses to the F1 and V antigen components of the fusion protein were also examined. Both F1 and the V antigen have been shown by other workers to be protective and now the group at USAMRMC has shown that rF1 and rV are the best candidates identified to date for a new plague vaccine.

Again, this fusion protein has a histidine tag, which will need to be removed prior to its use in humans.

DR. JEFFREY PULLEN

Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Yersinia pestis* (abstract 18)

This study attempts to identify important B and T cell epitopes within both the F1 and V antigens, however only B cells were addressed in the presentation. Identifying the functional epitopes in these proteins is important both to an understanding of the protective mechanisms stimulated by these two immunogens, and for assessing the potential of using synthetic peptides rather than entire recombinant proteins in a vaccine. This study is an important part of fulfilling the long-term objective of developing a useful vaccine. However, the usefulness of the current approach should be carefully reconsidered.

The use of short peptides to generate antibodies without conjugation to carrier molecules has, in general, not been very successful. Although it is sometimes possible to generate antibodies against short peptides, it is unlikely that the response will be protective without some T cell involvement. The investigators' initial experiments showed that peptides generated from the region of the protein known to be antigenic failed to generate a protective response despite generating significant antibody production. These results should have alerted them to the problems inherent in this approach. Instead, the investigators expanded their studies in response to these findings by making and testing additional peptides covering the whole of V antigen and F1 protein. This was a lot of work, using a lot of mice, that generated very little useful information. A simpler and more direct approach to begin mapping the reactive epitopes in these immunogens is to screen the overlapping peptides *in vitro* using antisera from animals or humans that have either had infections with *Y. pestis* or been immunized with native F1 and/or V antigen. Another concern is that in the future goals, it was stated that the response to the peptides, rather than to the native antigen will be tested to better determine the response. However, since the goal is to get protective antibodies, it seems that the response to native antigen, which is what the animals will see in an actual infection, is what should be measured.

It is also important for the investigators to determine the nature of a protective immune response to *Y. pestis* infection before restricting their focus and undertaking such labor-intensive studies to define only B cell epitopes. Antibody reactivity does not assure protection, and with some pathogens high antibody titers have even been correlated with disease progression. In addition, non-F1 antigens may evoke a

COMBINED RECOMMENDATIONS AND CONCLUSIONS

The USAMRMC's program to develop a new subunit vaccine for pneumonic plague has been very productive and has made significant advances towards this objective. The leader and research team are highly skilled, competent investigators and, with continued support, it is anticipated that a new vaccine for human trials is only a few years away. The investigators have used very effective immunization and challenge protocols to test immunogens in both mice and nonhuman primates for protection against plague following either parenteral or aerosol exposures to *Yersinia pestis*. Having the facilities to safely execute aerosol transmission studies is a critical component of this program. The team has confirmed and extended the data supporting the potential for both recombinant F1 and V antigens to afford significant protection. The work using the F1-V antigen fusion protein is exciting and represents a significant advance made by this team.

The team has examined numerous other antigens for identifying additional protective immunogens, especially for challenge with strains of *Y. pestis* lacking the F1 antigen. For such isolates, the V antigen and possibly YopD are the only useful candidates identified to date. The addition of one more antigen would likely solve the problem of non-responders, as well as strengthen the response in all individuals. The choice of antigens being tested for potential vaccine components appears somewhat random. These studies could be focused better by determining what proteins induce an immune response, thereby demonstrating which determinants are most likely being seen by the immune system. Although it is not possible to predict in advance which antigens are protective, the search could have been directed more towards antigens known to induce an antibody response in infected human patients and laboratory infected animals. Additional focus on the basis of immunity to plague challenge is also recommended. The investigators are also aware of the immunosuppressive effects of V antigen, and plan to examine the mechanisms involved. These types of studies should allow the team to "fine tune" the V antigen to increase its efficacy and safety as a vaccine component.

The development of *in vitro* correlates of immunity should be a high priority of the program and is currently the weakest area of the future plans. As discussed with the investigators, the assumption that protection is solely mediated by antibody has potential difficulties. Before continuing studies to determine important B cell epitopes, the role of T cells needs to be addressed in collaboration with immunologists. There are standard methods, such as adoptive transfer, to determine if T cells protect against challenge with *Y. pestis*. There are also *in vitro* techniques to determine if T cells taken from an immunized animal proliferate in response to specific antigens. The studies using synthetic peptides have potential, but this work needs to be done with conjugated peptides. Alternatively, peptides could be attached to larger inert particles that could be taken up by B cells or macrophages that then present the antigen on class II MHC molecules on their surface. Epitope mapping of the F1 and V antigen peptides using immune sera from natural infections would have been an appropriate first step.

APPENDICES

APPENDIX 1: AGENDA

APPENDIX 2: ABSTRACTS

REVIEW OF PLAGUE RESEARCH PROGRAM

USAMRIID

15 FEBRUARY 1996

0815-0830 Welcome and introduction
COL David Franz, DVM, Ph.D.

0830-0900 Overview of Plague Program
COL Arthur M. Friedlander, M.D.

Treatment

0900-0930 Antibiotic treatment of experimental pneumonic plague
COL Russell Byrne, M.D.

Role of F1 Capsule in Pathogenesis and Immunity

0930-1000 Protective efficacy of active immunization with purified F1
from *Yersinia pestis* and an *Escherichia coli*
recombinant strain against lethal parenteral and respiratory
plague challenge
MAJ Gerard P. Andrews, Ph.D.

1000-1015 Coffee Break

1015-1100 F1 capsule is not a required virulence factor for the mouse or
non-human primate
Patricia L. Worsham, Ph.D.
M. Louise Pitt, Ph.D.
LTC Kelly J. Davis, DVM

Role of Non-F1 Proteins in Pathogenesis and Immunity

1100-1130 Studies on the role of the pigmentation locus in the
pathogenesis of *Y. pestis*
Patricia L. Worsham, Ph.D.

1130-1300 Lunch

1300-1320 Analysis of the role of pPst encoded genes in pathogenesis of infection by *Y. pestis*
Susan L. Welkos, Ph.D.

1320-1335 Plasminogen activator protease degrades proinflammatory cytokines
Allen Sample, Ph.D.

1335-1405 Cloning, expression, and protective efficacy of Yops and pH 6 antigen
MAJ Gerard Andrews, Ph.D.

1405-1420 Cloning, expression, and protective efficacy of V antigen
LTC George J. Anderson, Jr., Ph.D.

1420-1435 Cloning, expression, and protective efficacy of F1-V fusion protein
LTC George J. Anderson, Jr., Ph.D.

1435-1450 Determination of important B and T-lymphocyte epitopes in the F1 and V antigen proteins of *Y. pestis*
Jeffrey Pullen, Ph.D.

1450-1515 Overview of future plans
COL Arthur M. Friedlander, M.D.

Recombinant F1-V (rF1-V) Fusion Protein Protects against Lethal Wildtype *Yersinia pestis* in a Mouse Model

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Bacteriology Division, U.S. Army Medical Research Institute of Infectious Diseases, Ft. Detrick, MD.

The virulence of F1- strains and their occurrence in nature imply that F1 immunogen will not be sufficient for an optimal new plague vaccine. A fusion protein has the theoretical possibility of simplifying and reducing the cost of production of multiple antigens in addition to stabilizing the protein. For these reasons, we developed a fusion protein consisting of both the F1 and V antigens (1). The first fusion protein made consisted of F1 fused with residues 168-175 of the V antigen, a segment which previous studies suggested to contain a protective epitope. This fusion protein was used with the adjuvant alhydrogel (aluminum hydroxide) to immunize female Swiss Webster (Hsd:ND4) mice subcutaneously (s.c.) on days 0 and 30 followed by a s.c. or aerosol challenge with either the F1- C12 strain (LD₅₀ = 9.1 CFU, s.c.; LD₅₀ = 1.1 x 10⁵ CFU, aerosol route) or the wild-type F1+ CO92 (LD₅₀ = 1.9 CFU, s.c. route; 2.1 x 10⁴ CFU, aerosol route) strain of *Y. pestis*. Endotoxin had been removed from the preparations prior to immunization, so that this would not be a confounding factor.

When 18.5 µg of the F1-V168-275 fusion protein was used to immunize mice, there was 90% survival (9/10) when challenged s.c. with 63 LD₅₀ of the F1+ CO92 strain. The positive control was a group of mice immunized with 10 µg of rF1 which is equivalent to the F1 content of the F1-V168-275 protein. The rF1 control resulted in 100% (10/10) protection. The F1-ELISA IgG titers were the same (1:81920). All mice in alhydrogel control group died (0/9; MTD ± SD, 5.2 ± 1.0). When the F1-V168-275 immunized mice were challenged with 104 LD₅₀ by the aerosol route, protection was 80% (8/10; MTD ± SD, 20.3 ± 7.1) compared to 0% for the control group (0/10; MTD ± SD, 3.1 ± 0.3; 80-104 LD₅₀). The group immunized with rF1 resulted in 70% protection (7/10; MTD ± SD, 9.0 ± 1.0) when challenged with 80 LD₅₀. The addition of part of the V protein onto the F1 protein did not appear to effect its antigenicity.

The F1- strain, C12, was used to test the ability of the partial V portion of the F1-V168-275 protein to protect mice against a lethal challenge. Here 27 µg of the F1-V168-275 fusion protein was used, which is equivalent to 10 µg of the V protein

known to be protective. A s.c. challenge dose of 55 LD50 resulted in 30% survival (3/10, MTD \pm SD, 9.4 \pm 7.0). All of the controls died (0/10, MTD \pm SD, 10.8 \pm 4.8). While there was some protection, there was no increase in the MTD. There was a good V-ELISA antibody response to the F1-V168-275 (1:163840). In case this response was not sufficient, another group was immunized with 27 μ g, but with complete Freund's adjuvant (CFA). In this case, protection was only 20% (2/10, MTD \pm SD, 9.1 \pm 3.2), while 10 μ g of rV in CFA resulted in 100% protection. The V-ELISA titer when CFA was used was 1:1310720 for F1-V168-275 and rV. A 10-fold increase in the V-antibody titer did not have any effect on protection and the V-ELISA titer was not indicative of protection. When a group of F1-V168-275 mice were challenged with 95 LD50, C12, by the aerosol route, no mice survived (0/10, MTD \pm SD, 3.5 \pm 0.5). All of the alhydrogel control group died (0/10, MTD \pm SD, 3.4 \pm 0.5). In other experiments, rV itself gave 80-90% protection against an aerosol challenge.

These results demonstrated the feasibility of making a F1-V fusion protein. The efficacy of F1 was not altered by making a fusion protein. However, while the V168-275 protein portion of the fusion protein was antigenic, it was not immunogenic. This caused us to address the question as to whether the entire V protein could be fused to F1 and whether it would be immunogenic.

Using a fusion protein which combines the whole F1 and the whole V protein (rF1-V) to immunize mice on days 0 and 30 increased the protection afforded by the V portion of the fusion protein. When 13.6 μ g of rF1-V was used to immunize mice, there was 100% (10/10) protection against a s.c. challenge of 57 LD50 and 90% (9/10) protection against 1.1 \times 10⁶ LD50 C12 strain. Ten micrograms (10 μ g) of rV also gave 90% (9/10) protection against 1.1 \times 10⁶ LD50, C12 strain. All of the alhydrogel control group died (0/10, MTD \pm SD, 6.0 \pm 0.0). The rF1-V protein also offered protection against an aerosol challenge. The same immunization schedule resulted in 100% (10/10) when mice were challenge with 546-636 LD50, C12 strain on day 73 postimmunization. When mice immunized with the rF1-V fusion protein were challenged with 762 LD50 of the F1+, CO92 strain by the aerosol route, 100% (10/10) of the mice survived. The F1-V fusion protein was able to protect mice from a significant aerosol challenge from either a F1+ or F1-lethal strain of Y. pestis. This protection is better than the protection afforded by the current Plague Vaccine USP. When mice which were immunized on day 0 and 30 with 0.2 ml of the current vaccine and challenge by the aerosol route on day 73 postimmunization with 546-636 LD50, C12 strain, all of the mice died (0/8, MTD \pm SD, 3.3 \pm 0.5). The V-ELISA titer to the Plague

Vaccine USP was <1:640.

The recombinant rF1-V fusion protein was produced in *E. coli* and contained a polyhistidine tag which aids in the purification of the fusion protein. While this protein has been shown to be highly efficacious in the mouse model, it remains to be seen whether this level of protection will be seen in the non-human primate model. Further, the regulatory issue of whether a histidine tagged protein will be acceptable to the Food and Drug Administration needs to be resolved.

1. Heath, D.G., G.W. Anderson, Jr., J. M. Mauro, S.L. Welkos, and A.M. Friedlander. Protection against experimental bubonic and pneumonic plague by a recombinant capsular F1-V antigen fusion protein vaccine. manuscript submitted.
2. Brubaker, R.R., A.K. Sample, D.Z. Yu, R.J. Zahorchak, P. C. Hu, and J.M. Fowler. 1987. Proteolysis of V antigen from *Yersinia pestis*. *Microbial Pathogenesis*. 2:49-62.

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